

Le déclenchement de la véraison

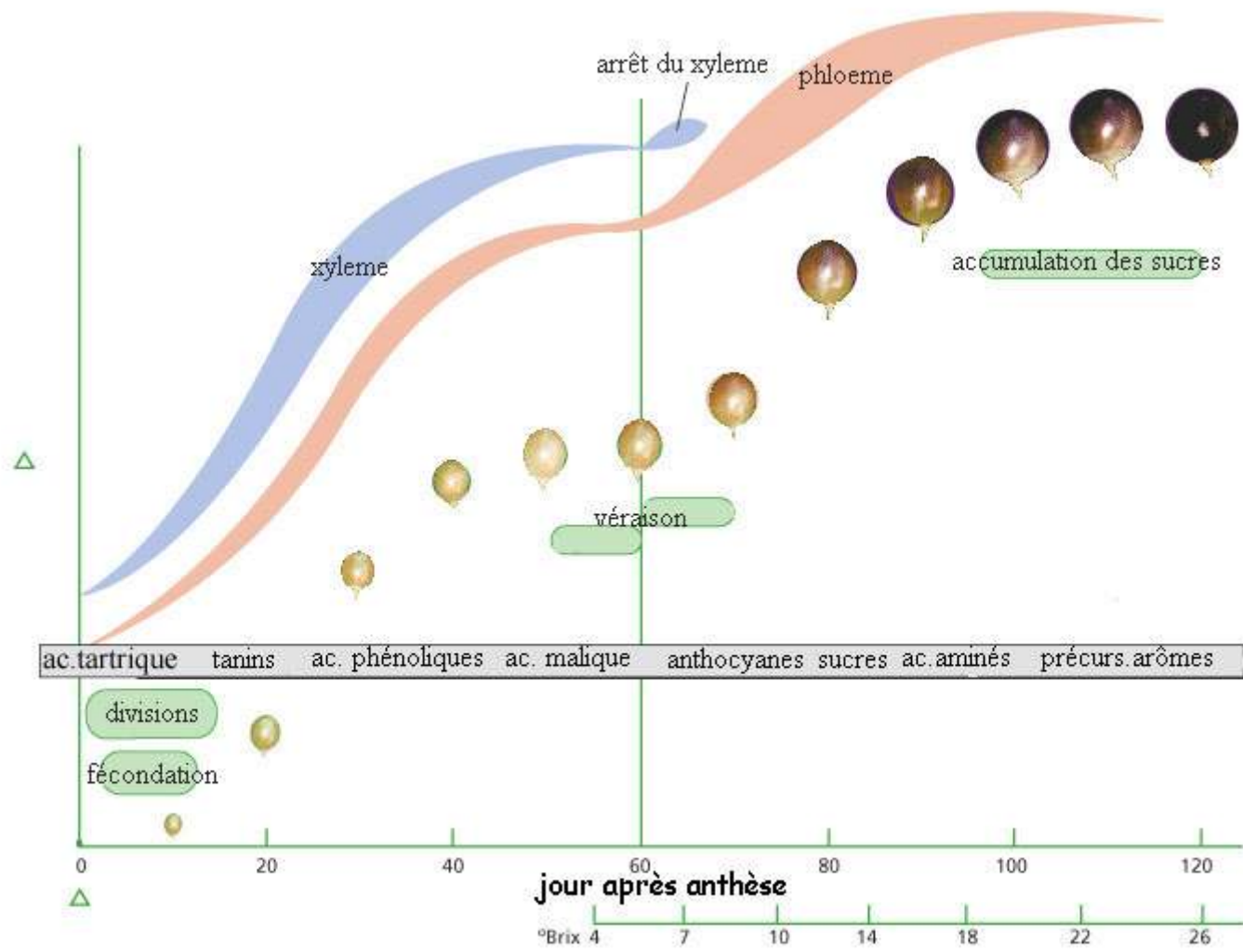
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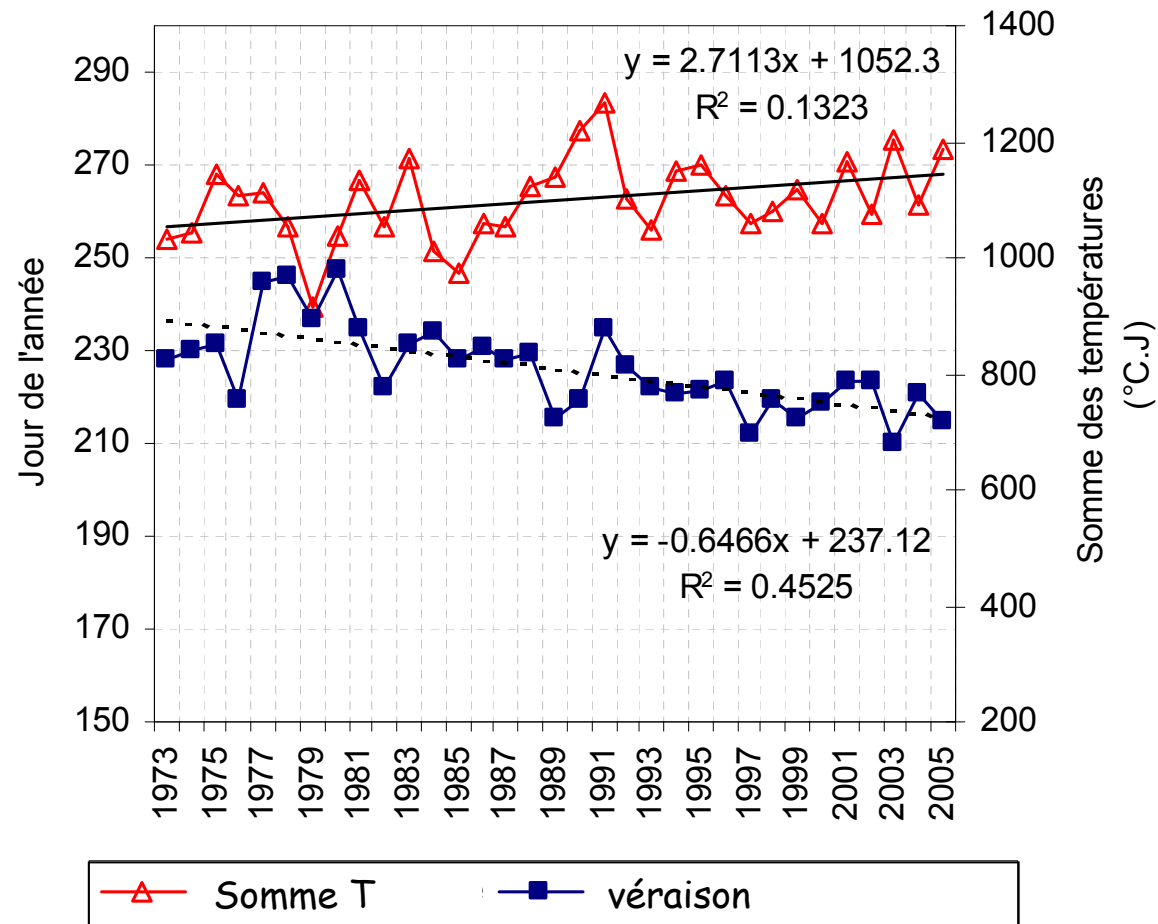
10 mars 2006



Les phases de développement de la baie de raisin

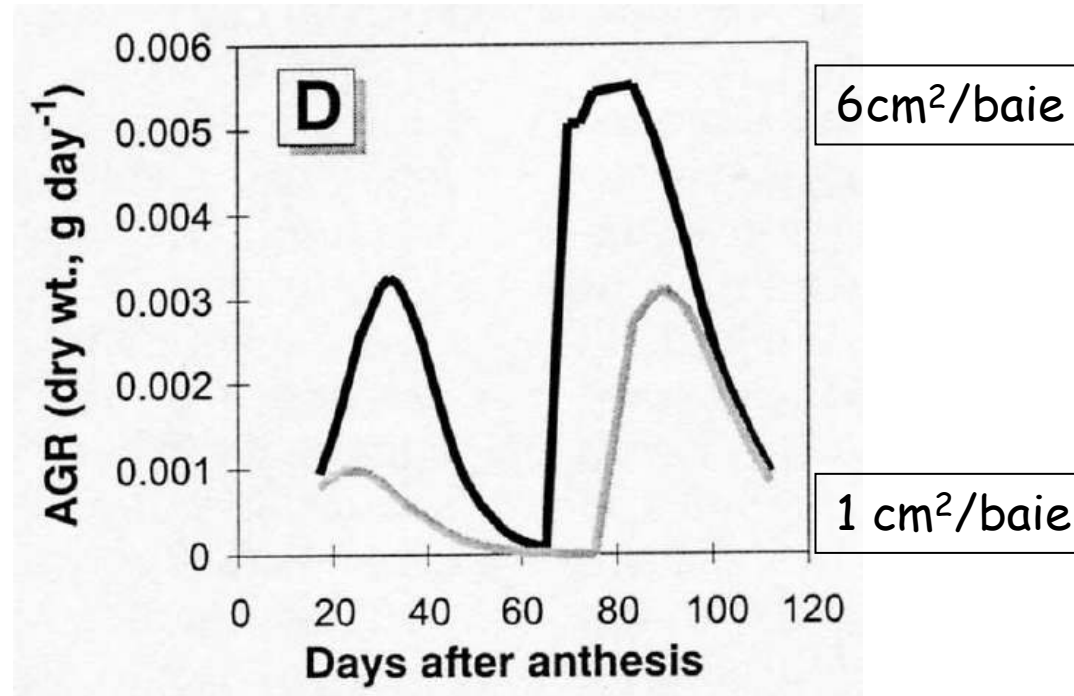


La date de la véraison dépend de la température



Evolution des dates de mi-véraison
du Cabernet Sauvignon et du Merlot
en Gironde.

Une photosynthèse limitante retarde la véraison



Vitesse de croissance d'une baie de raisin en fonction du rapport feuille/fruit (Ollat et Gaudillere, 2002)

Le déclenchement de la véraison est un phénomène localisé à la baie



Structure de la baie de raisin

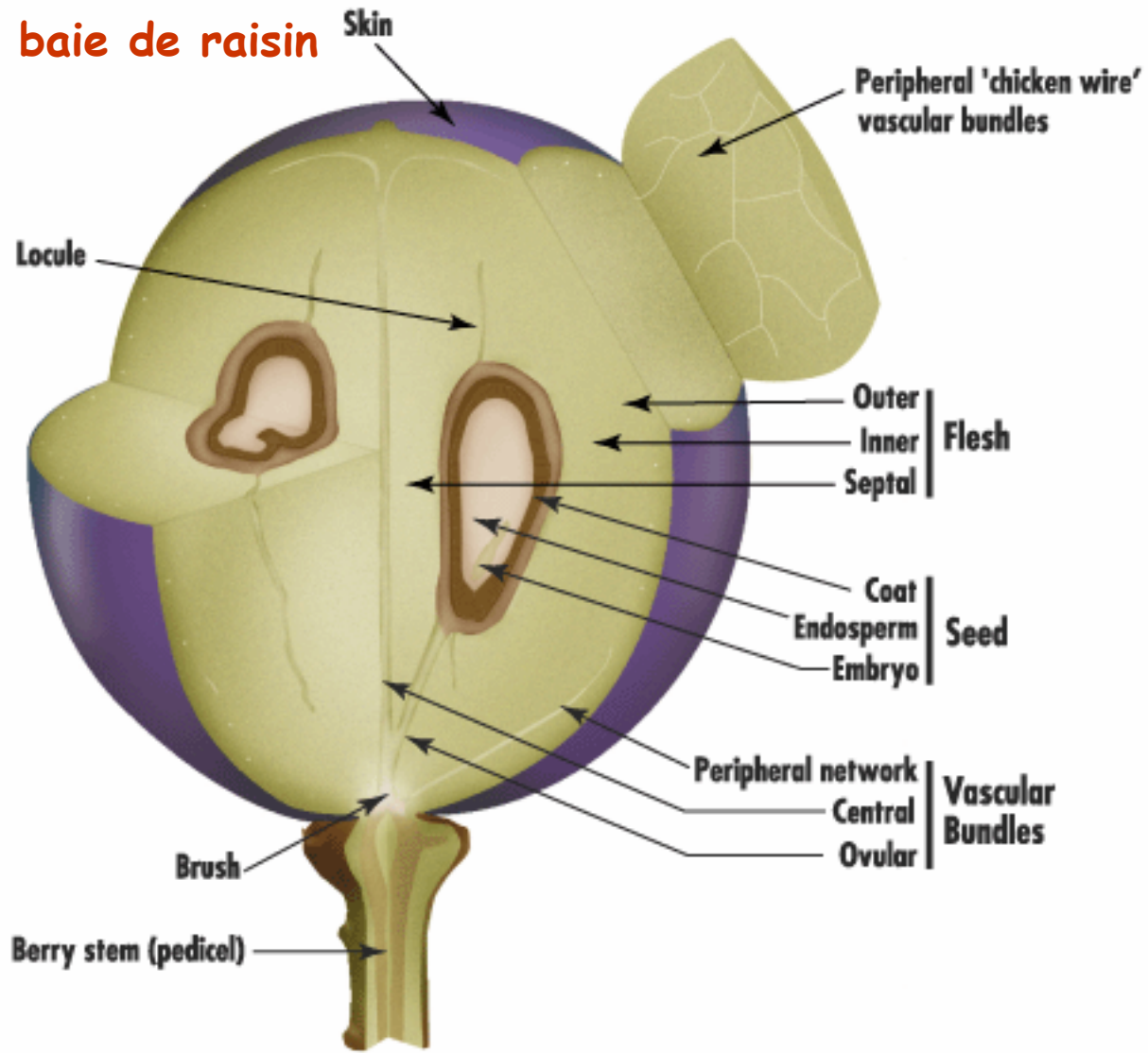


Figure 1: Structure of a ripe grape berry partially sectioned on the long and central axis to show internal parts. Illustration by Jordan Koutroumanidis, Winetitles.

1) Ramollissement des structures pariétales

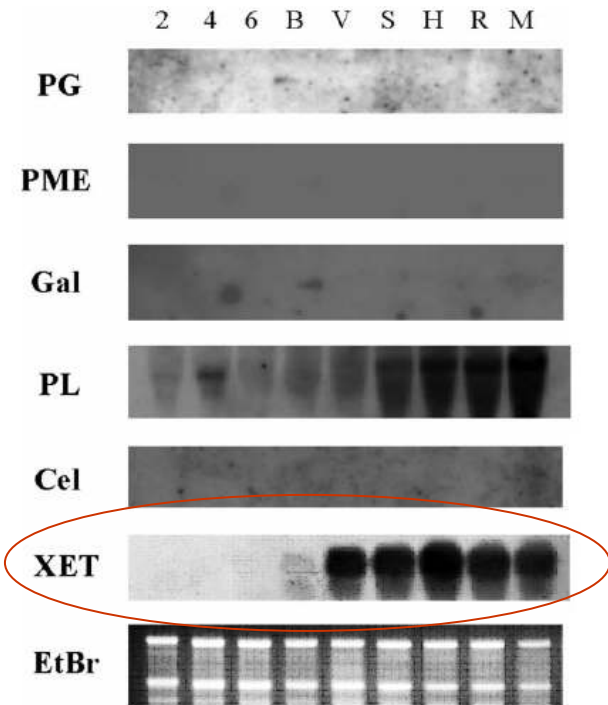


Fig. 1. Northern blot analysis of RNA from *Kyoho* berries taken at several stages of development. Total RNAs (20 µg) were separated by electrophoresis and then transferred onto nylon membranes. The membranes were hybridized with DIG-labeled cDNAs for cell wall degradation-related enzymes, which were obtained from a subtractive library (PG, PME, GAL, PL, Cel, and XET). The numerals indicate weeks after flowering, and B, V, S, H, R, and M indicate just before véraison, véraison, beginning color, half-colored, fully colored reddish purple, and fully colored black (mature), respectively.

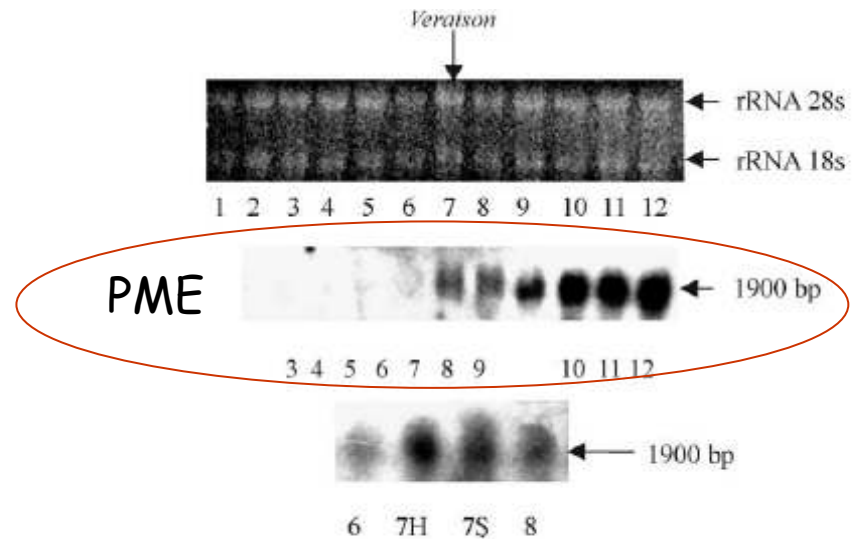


Fig. 1. Upper panel, the development of Ugni blanc berries. A, average berry fresh weight (□, 1996 and ●, 1997). B, pH (□, 1996 and ○, 1997) and °Brix (■, 1996 and ●, 1997) of the resulting must. Lower panel, expression pattern of mRNA coding for *pme1*. Fifteen micrograms of total RNA were separated on 1.2% denaturing agarose-formaldehyde gel electrophoresis and transferred onto a nylon membrane. C, total RNA blots were stained with ethidium bromide as a loading control; northern blots were hybridized with the *pme1* probe (³²P labelled cDNA); D, total RNA from harvest 1997, week 7 (*véraison*) corresponding to a mix of ~50% hard and soft berries from *véraison* stage; E, northern blot differentiating hard (7H) and soft (7S) berries from the *véraison* stage (week 7).

1) Ramollissement des structures pariétales

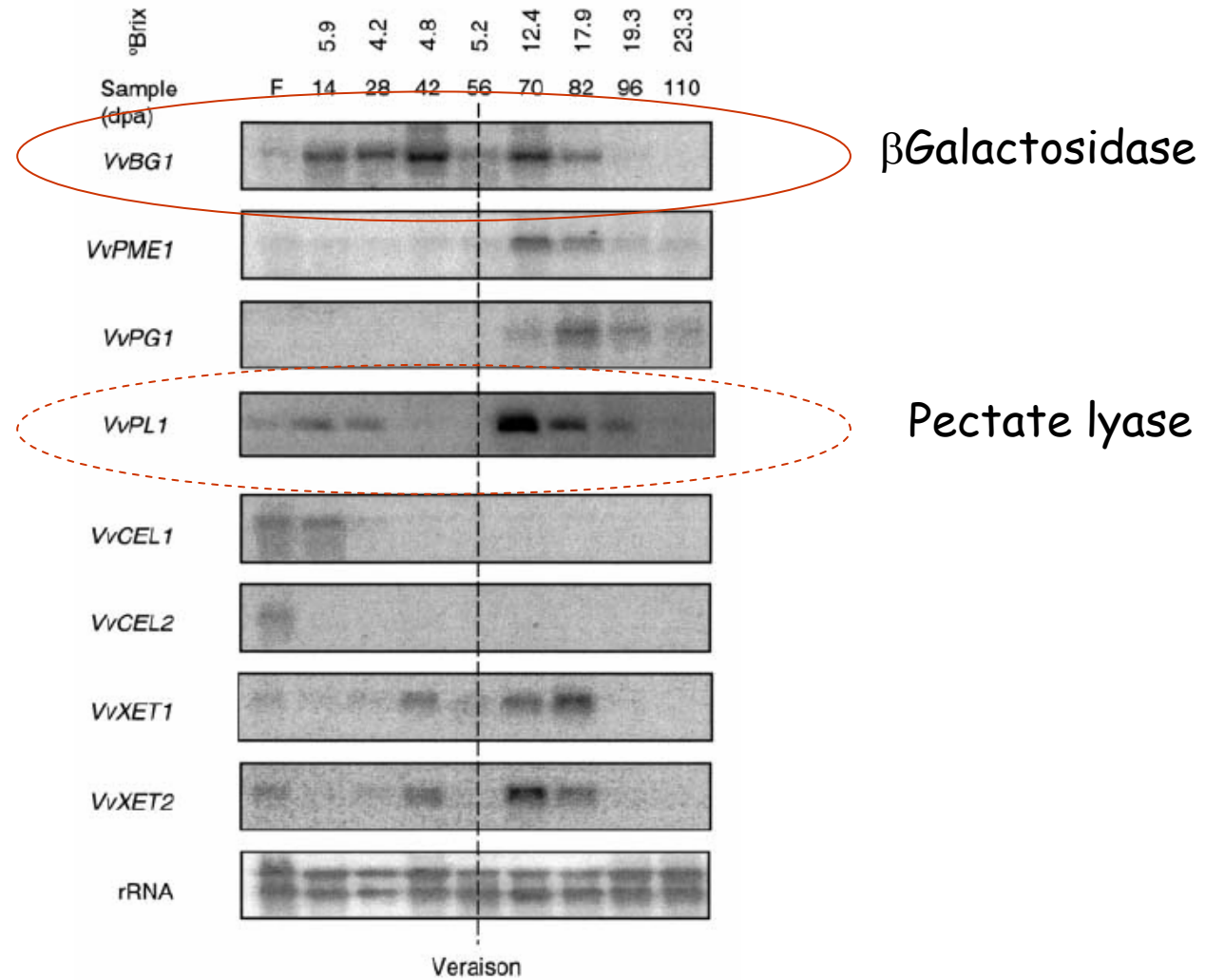
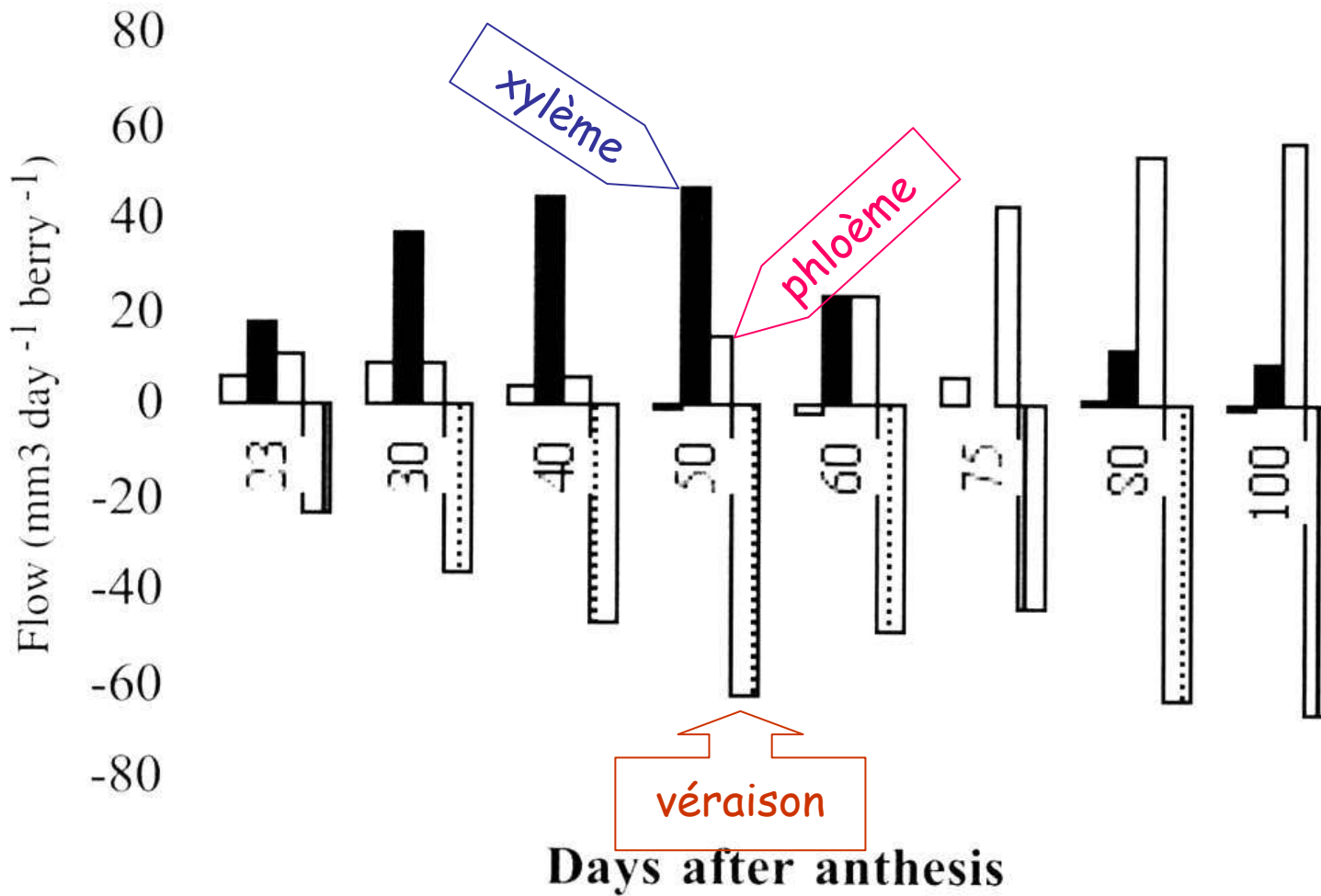


Fig. 4 Northern analyses of RNA extracted from Shiraz grape flowers (*F*) and developing Shiraz berries. °Brix and time of sampling, from flowering through to ripeness (110 dpa) are indicated above each lane. Total RNA (2.8 µg) was loaded in each lane. The membrane originally probed with *VvBG1* was re-probed with a ribosomal RNA to ensure similar loadings in each lane

2) Arrêt du fonctionnement du xylème,
stimulation du transport phloémien



□ net flow ■ xylem flow ◻ phloemic flow ◻ transpiration

3) Stockage des sucres et désacidification

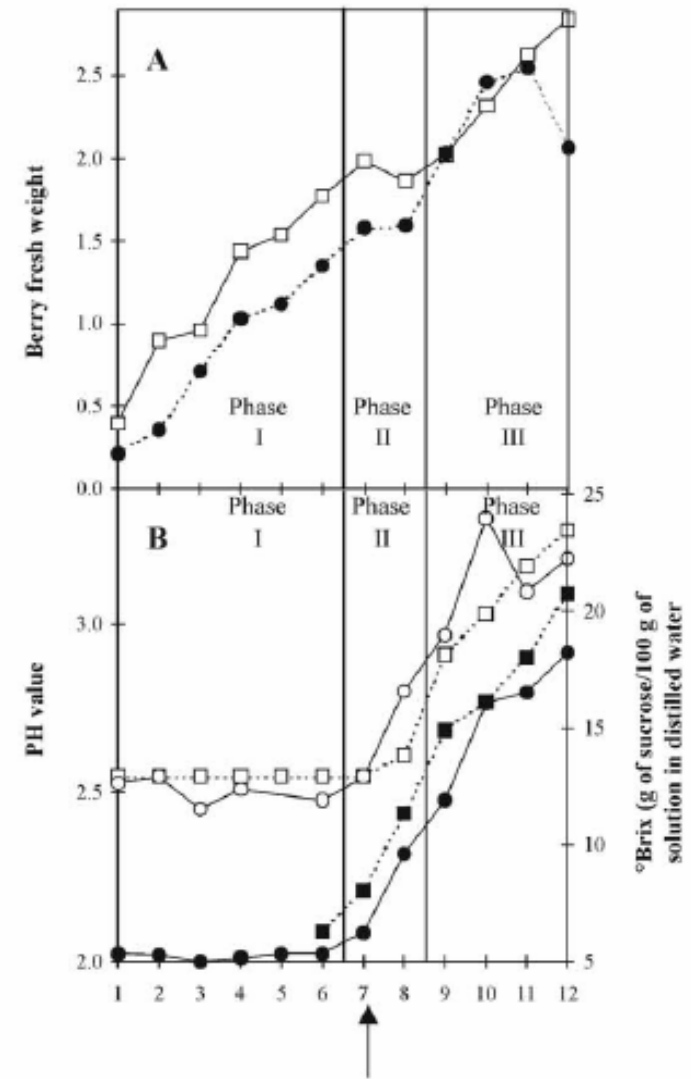
❖ Arrêt de la synthèse des principaux acides organiques

malique } dans la pulpe
tartrique }
oxalique } dans la pellicule

❖ Changement de destination des sucres
stockage vacuolaire du glucose et fructose

❖ Stockage d'acides aminés, arginine, proline

❖ Décompartimentation et dégradation de l'acide malique et désacidification de la pulpe



4) Synthèse massive de protéines pariétales et cytosoliques

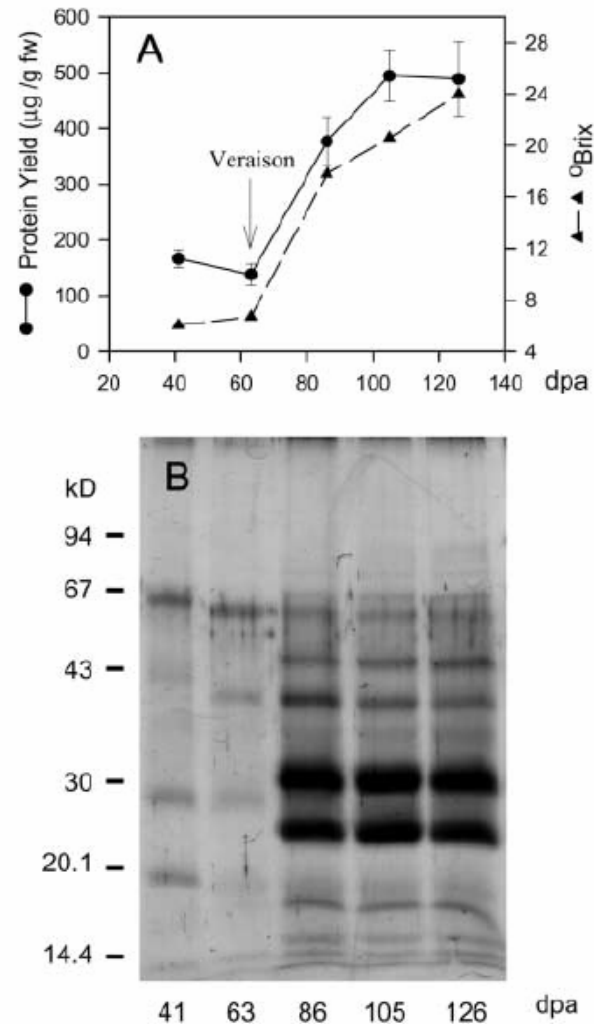


Fig. 1 a Yields and profiles of proteins extracted from developing Gordo grape (*Vitis vinifera*) berries (41–126 dpa). b Profiles of the extracted proteins were resolved by SDS-PAGE. Each lane on the gel contained protein extracted from approximately 42 mg fresh weight of tissue. The position of molecular mass markers is indicated on the left

4) Expression des gènes des protéines GRIP

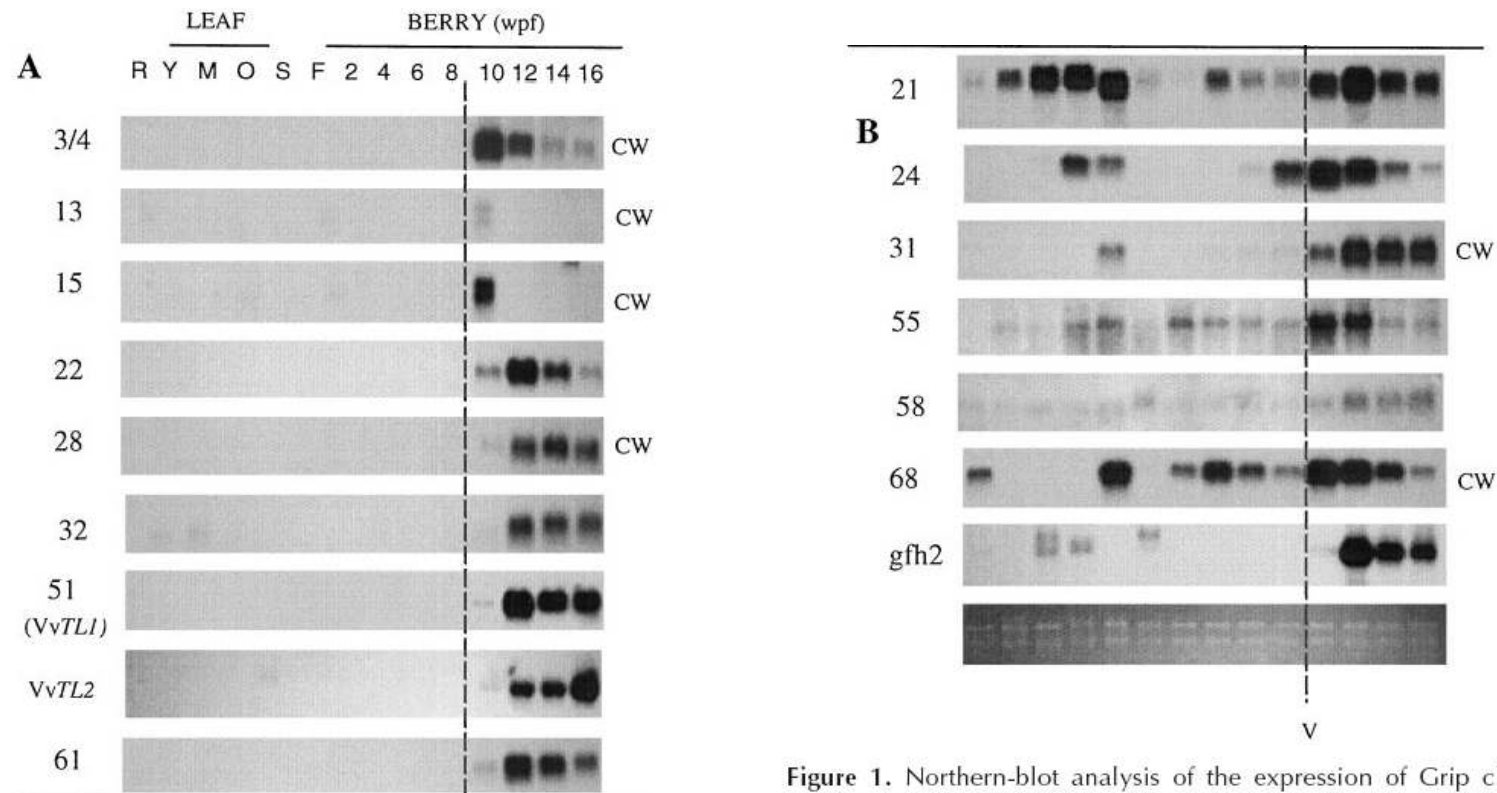
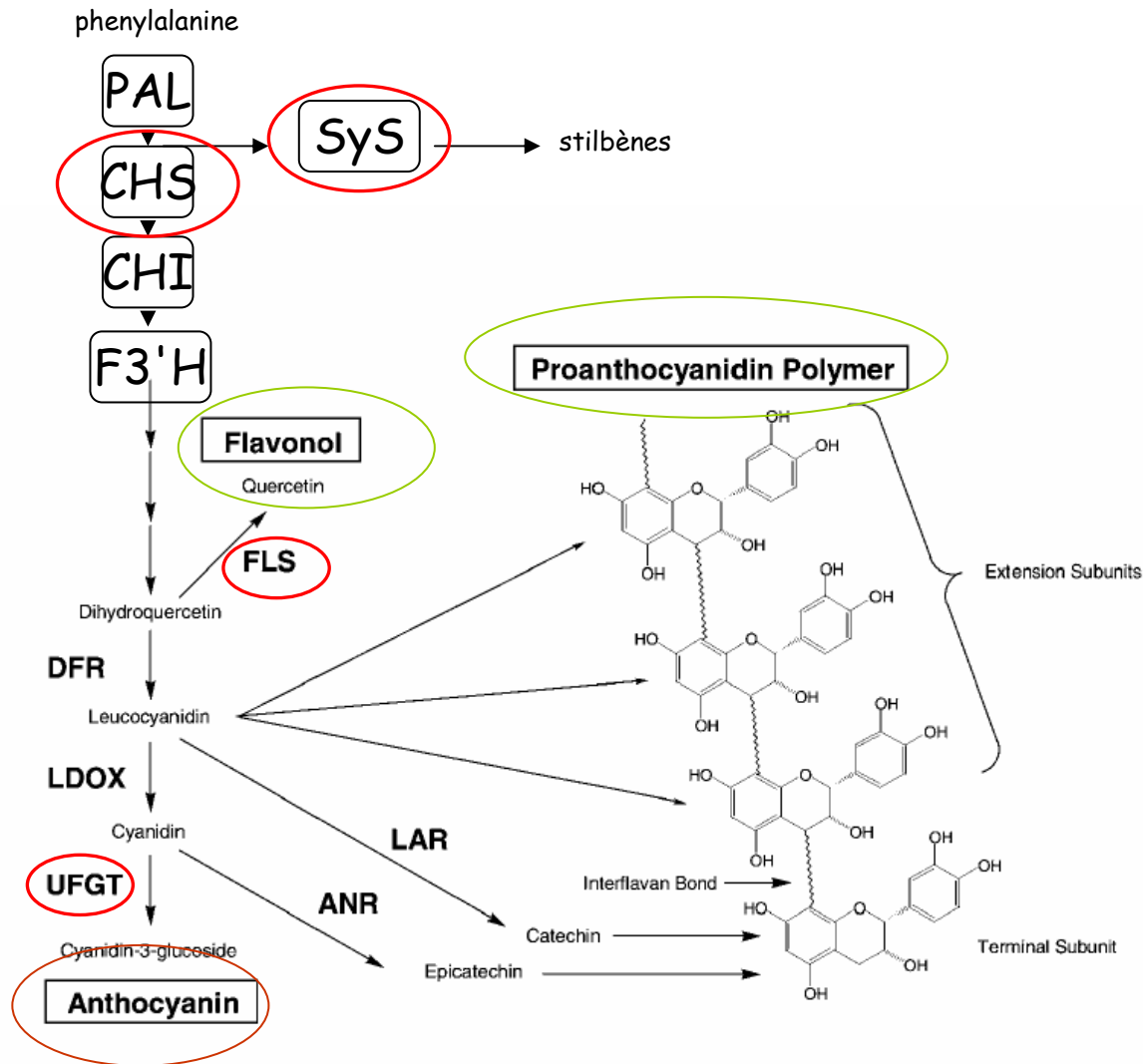


Figure 1. Northern-blot analysis of the expression of Grip clones (and the gfh2 and VvTL2 clones) in various grape tissues. RNA from root (R), young leaf (Y), mid leaf (M), old leaf (O), seed from berries 4 wvf (S), flower (F), and a series of samples taken from developing berries at two weekly intervals (commencing at 2 wvf) were probed with the cDNAs as indicated. The dashed line indicates véraison. A, Genes exhibiting berry- and ripening-specific expression. B, Genes with up-regulated expression during ripening but also expressed in other tissues. CW, Clones encoding putative cell wall proteins. The bottom panel is a photograph of an ethidium-bromide-stained gel to show the intactness and relative loadings of the RNA samples used in the northern analysis.

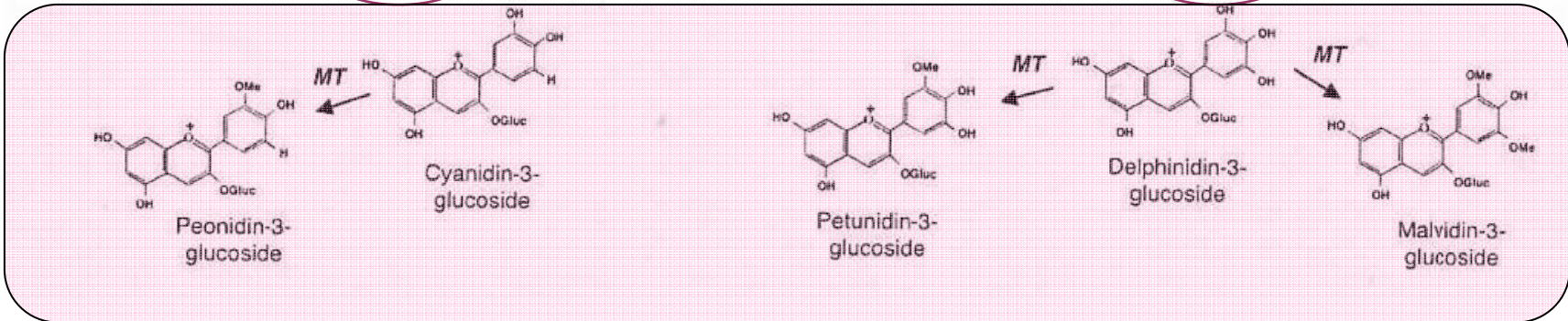
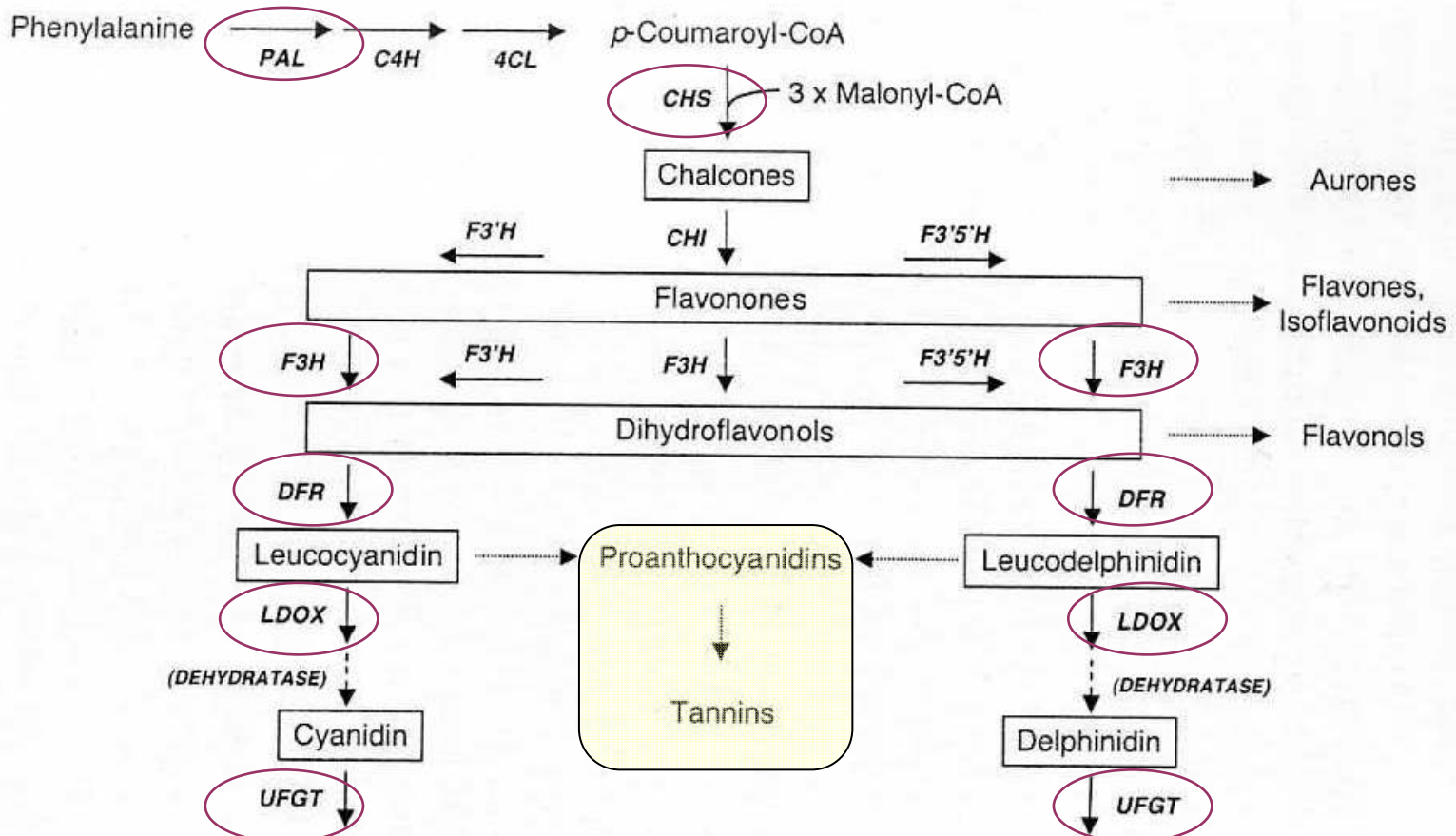
5) Flavonoids biosynthesis in Grapes



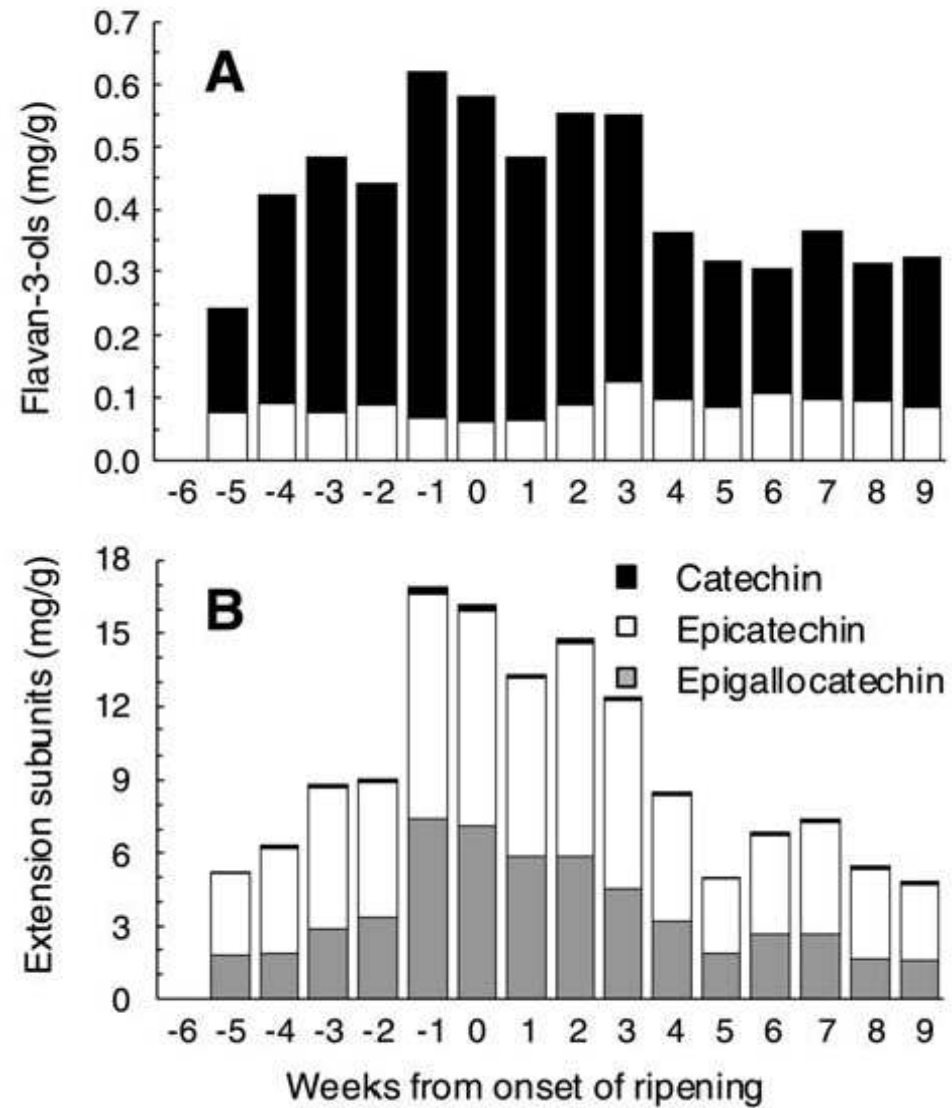
Proanthocyanidin Biosynthesis in Grapes

Figure 1. Scheme of the flavonoid pathway leading to synthesis of anthocyanins and PA polymers. Leucocyanidin is a precursor to synthesis of anthocyanins such as cyanidin-3-glucoside but also contributes the extension subunits of the PA polymer. LAR catalyzes reduction of leucocyanidin to catechin and ANR (*BANYULS*) catalyzes reduction of cyanidin to epicatechin, both of which can form the terminal unit of the PA polymer. FLS, Flavonol synthase.

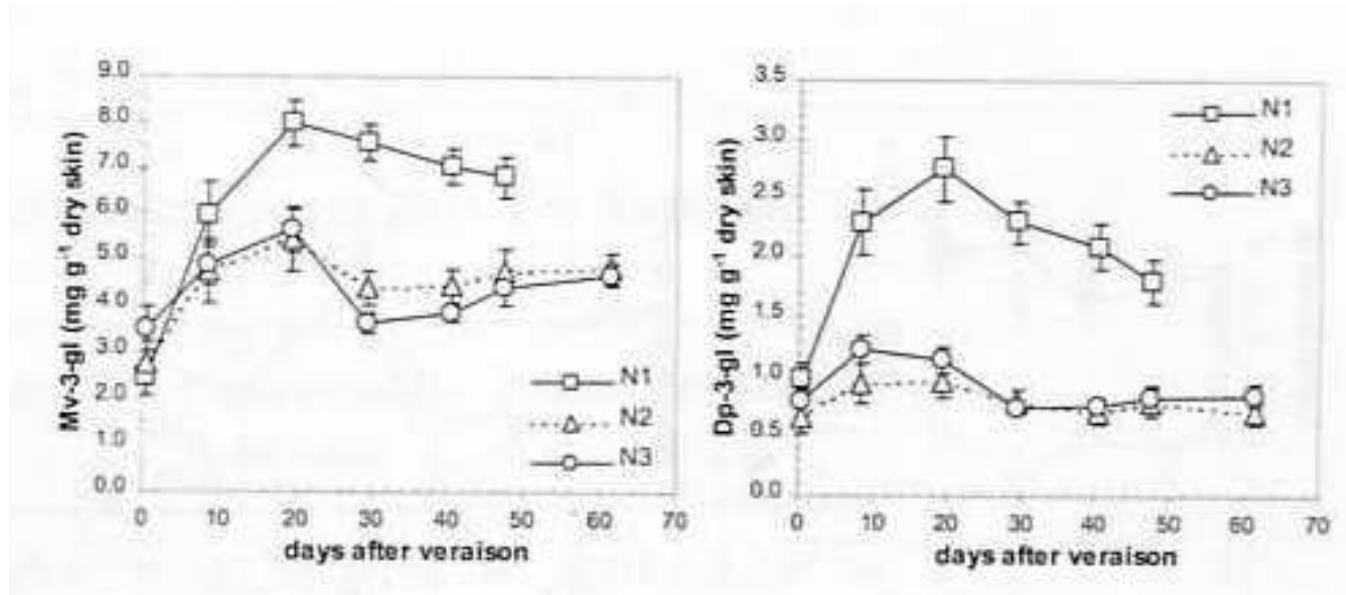
Métabolisme des flavonoïdes



Accumulation of proanthocyanidins in berry skin



Synthèse des anthocyanes post véraison



Perte de la capacité de produire des stilbènes

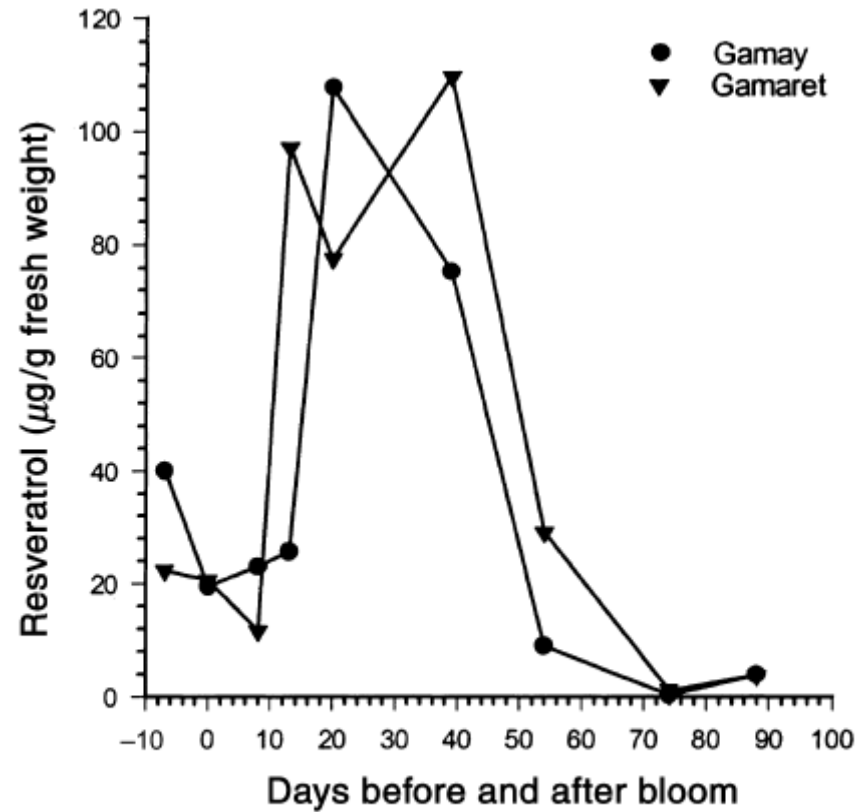


Fig. 4 Concentration of resveratrol in grape berries of Gamay and Gamaret at stages 1-9, 24 h after UV irradiation

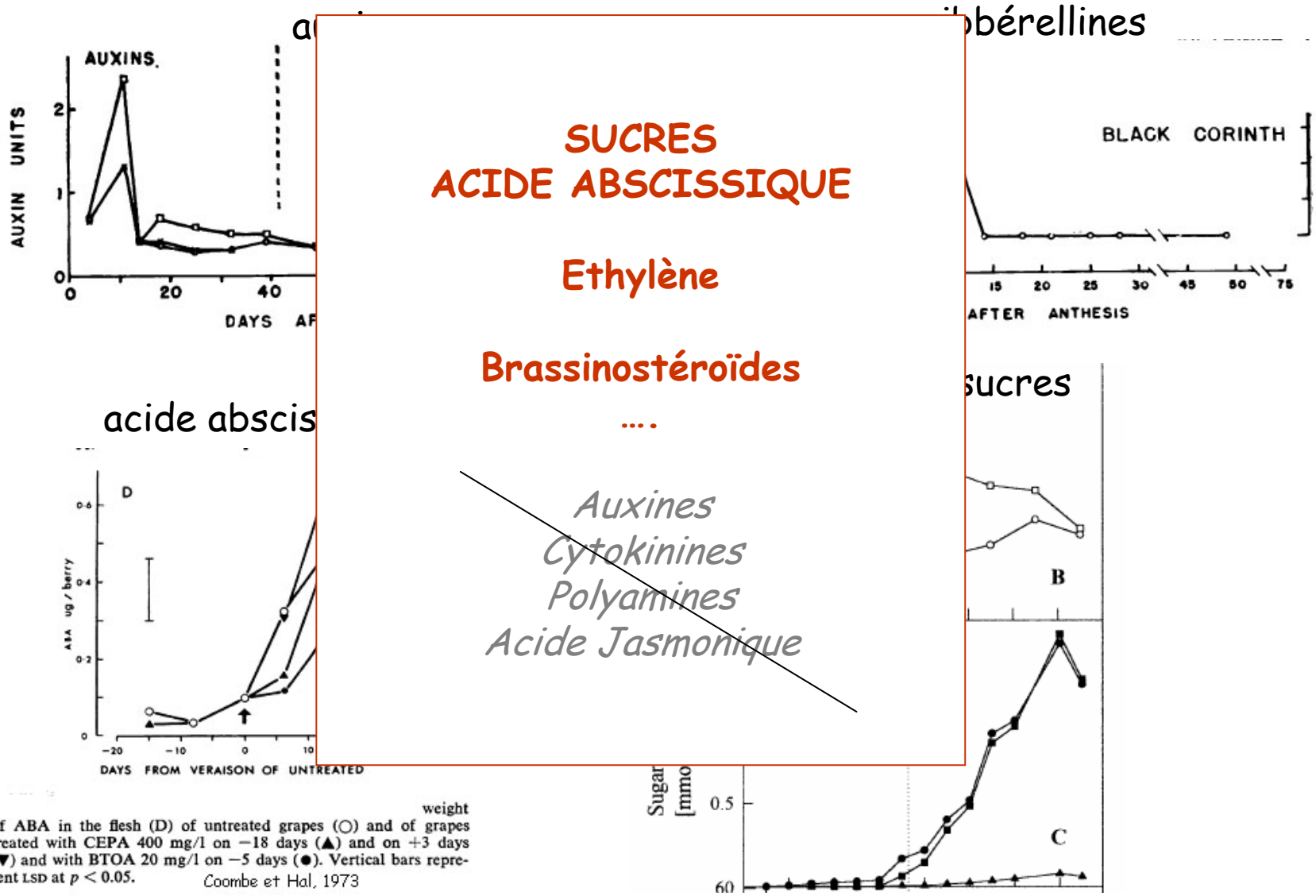
6) Nature des signaux impliqués au moment de la véraison

Rechercher signaux qui agissent sur des voies métaboliques modifiées à la véraison:

- synthèse des anthocyanes
- transport des sucres
- induction de l'ADH
- désacidification

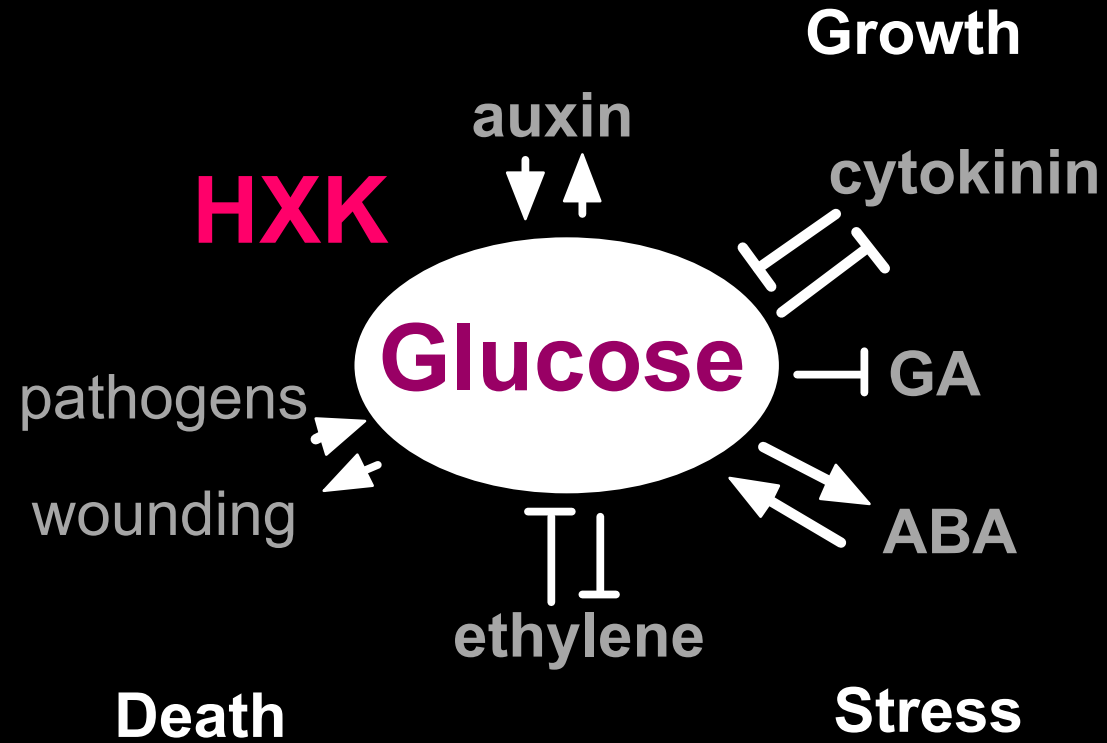
Identifier les signaux qui varient à la véraison:

6) Nature des signaux impliqués au moment de la véraison



Il faut rechercher une régulation en réseau

Glucose is a central signaling molecule



Arabidopsis

7) en face des signaux il y a des récepteurs

- ❖ Régulation de l'expression des gènes (promoteurs...)
- ❖ Régulation de la production de facteurs de transcription: MYB ...
- ❖ Protéines régulatrices: VvMSA protéine induite par les sucres,

Nécessité d'intégration par la bioinformatique
et la modélisation aux différents niveaux

génomique
transcriptome
protéome
métabolome
plante entière

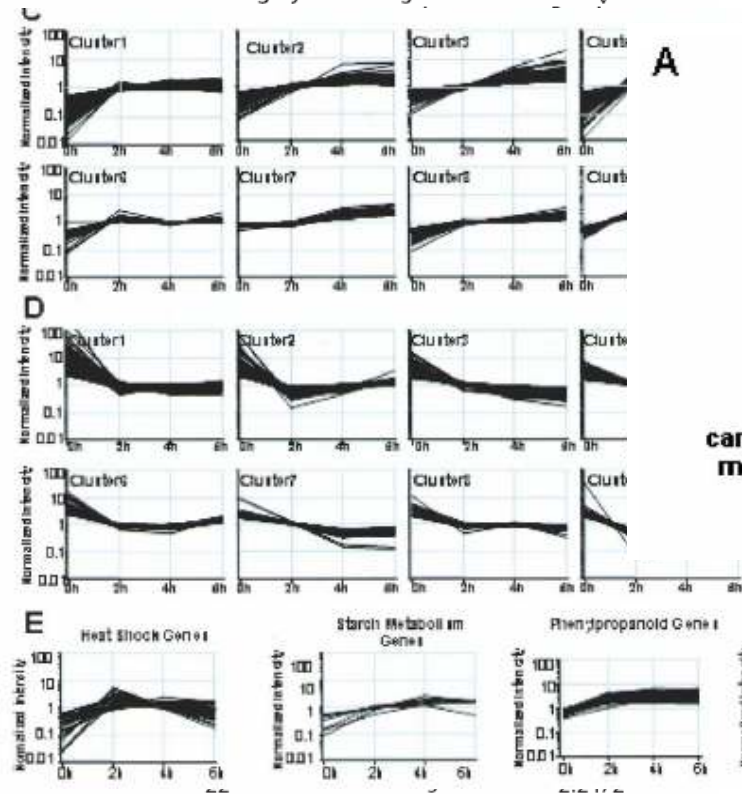
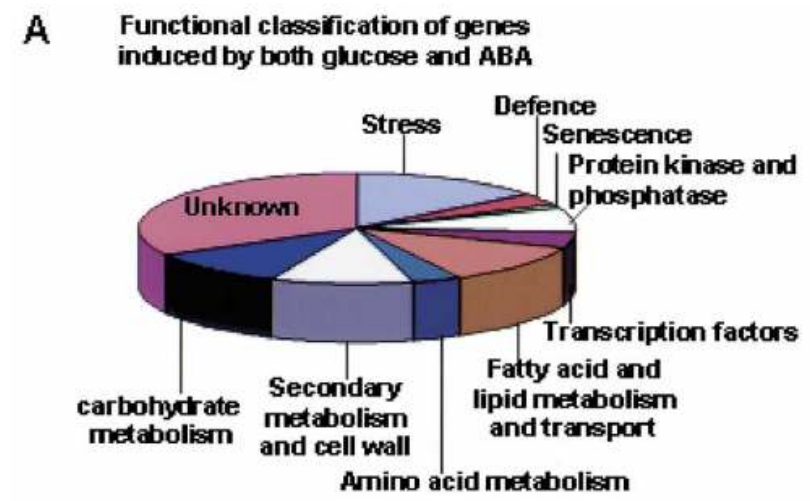
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Recherche de séquences promotrices inconnues avec les outils de la génomique

Establishing glucose- and ABA-regulated transcription networks in *Arabidopsis* by microarray analysis and promoter classification using a Relevance Vector Machine

Table 1. Functional categorization of glucose- and ABA-responsive genes

Category	Genes in category	Regulated	P_v	
Glucose up-regulated	Cluster1	Cluster2	Cluster3	Cluster4
Sulfur assimilation	Cluster5	Cluster7	Cluster8	Cluster9
Translation	Cluster10	Cluster11	Cluster12	Cluster13
Abiotic stress	Cluster14	Cluster15	Cluster16	Cluster17
Nucleotide synthesis	Cluster18	Cluster19	Cluster20	Cluster21
Protein targeting	Cluster22	Cluster23	Cluster24	Cluster25
Secondary metabolism methyl transfe	Cluster26	Cluster27	Cluster28	Cluster29
Lipid transfer proteins	Cluster30	Cluster31	Cluster32	Cluster33
Secondary metabolism flavonoid synt	Cluster34	Cluster35	Cluster36	Cluster37
Nucleotide sugar transferases	Cluster38	Cluster39	Cluster40	Cluster41
Ribosomal proteins	Cluster42	Cluster43	Cluster44	Cluster45
Glucose down-regulated	Cluster46	Cluster47	Cluster48	Cluster49
Amino acid degradation	Cluster50	Cluster51	Cluster52	Cluster53
Trehalose metabolism	Cluster54	Cluster55	Cluster56	Cluster57
Glutaredoxins	Cluster58	Cluster59	Cluster60	Cluster61
Gluconeogenesis	Cluster62	Cluster63	Cluster64	Cluster65
Storage proteins	Cluster66	Cluster67	Cluster68	Cluster69
Ethylene synthesis	Cluster70	Cluster71	Cluster72	Cluster73
Pentose phosphate pathway	Cluster74	Cluster75	Cluster76	Cluster77
Lipid degradation	Cluster78	Cluster79	Cluster80	Cluster81
Ubiquitin conjugation	Cluster82	Cluster83	Cluster84	Cluster85
Light-mediated signaling	Cluster86	Cluster87	Cluster88	Cluster89
ABA up-regulated	Cluster90	Cluster91	Cluster92	Cluster93
Abscisic acid metabolism	Cluster94	Cluster95	Cluster96	Cluster97
Secondary metabolism	Cluster98	Cluster99	Cluster100	Cluster101
General carbohydrate degradation	Cluster102	Cluster103	Cluster104	Cluster105
Storage and lipid transfer proteins	Cluster106	Cluster107	Cluster108	Cluster109
Lipid metabolism	Cluster110	Cluster111	Cluster112	Cluster113
Salicylic acid metabolism	Cluster114	Cluster115	Cluster116	Cluster117
P450 and degradation enzymes	Cluster118	Cluster119	Cluster120	Cluster121
Trehalose metabolism	Cluster122	Cluster123	Cluster124	Cluster125



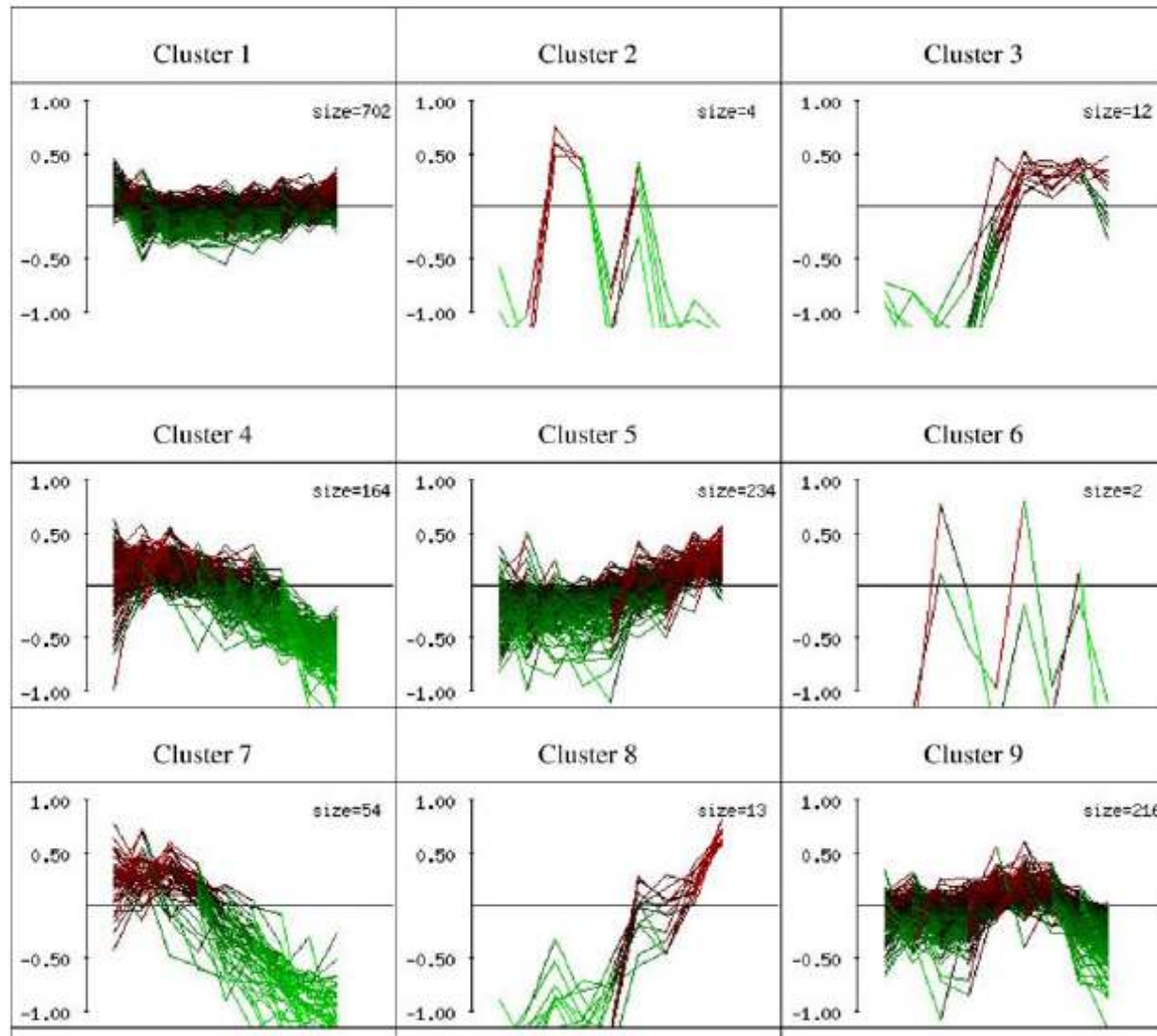
P -values of P_v corrected for multiple comparisons using the Bonferroni method in each regulated group are indicated with an asterisk.
 Glucose up-regulated, 43 functional categories, $-\log_{10} P_v = 3.445$.
 Glucose down-regulated, 19 functional categories, $-\log_{10} P_v = 3.365$.
 ABA up-regulated, 113 functional categories, $-\log_{10} P_v = 3.354$.

Utilisation de méthodes statistiques adaptées, réseaux de neurone, statistique Bayésienne...

(Li et al, Genome research, 2006)

Stratégie pour retrouver des régulateur de l'expression des gènes à la véraison

Identification des gènes co-régulés à l'aide des μ réseaux



Conclusions

Les hormones ne sont que des éléments ponctuels dans un schéma complexe de régulation qui fait intervenir:

- l'expression de nombreux gènes
- la synthèse des protéines codées par ces gènes
- l'activité de ces protéines
- L'intégration des processus

Les pratiques viticoles permettent le plus souvent d'assurer de façon satisfaisante la réalisation de cette phase de développement du raisin.

Le progrès des connaissances sur la véraison participe à l'amélioration de la gestion raisonnée de la vigne pour atteindre un état de maturation adapté à l'objectif technologique.



10 mars 2006

