



*VINELINK INTERNATIONAL*  
*www.liendelavigne.org*

**ASSEMBLEE GENERALE 2017**  
*2017 General Assembly*

**NOUVEAUX OUTILS POUR LE SUIVI DE**  
**LA QUALITE DES RAISINS :**

**Capteurs, analyse des données, outils**  
**d'aide à la décision**

*New tools for monitoring*  
*grapes quality : sensors, data*  
*analysis, decision*

# Hyperspectral imaging for viticultural applications

Javier Tardaguila





**10-year-experience in precision viticulture**

**New non-invasive sensing technologies**

**Vineyard monitoring / Phenotyping**

# Vineyard monitoring / Phenotyping



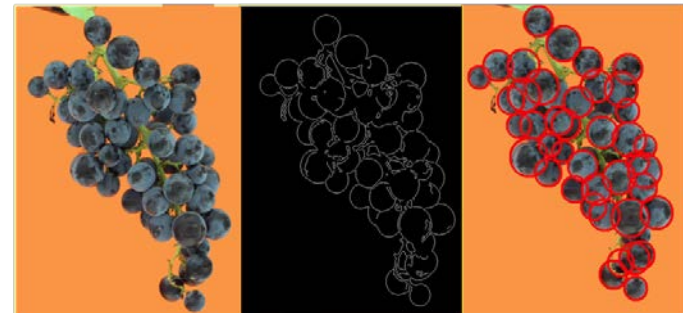
Water status



vitisFlower Apps



Grape composition



Yield components

# Televitis mobile lab

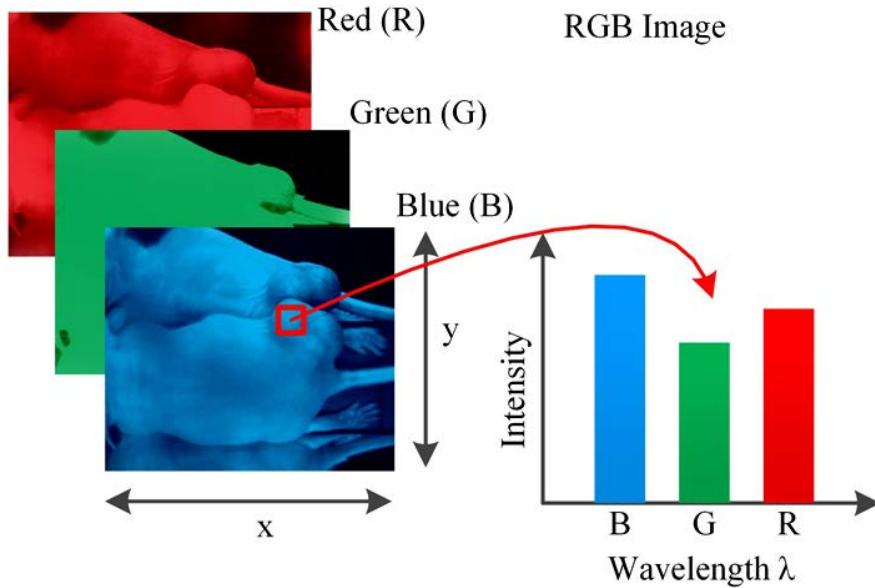


# Robotics in viticulture

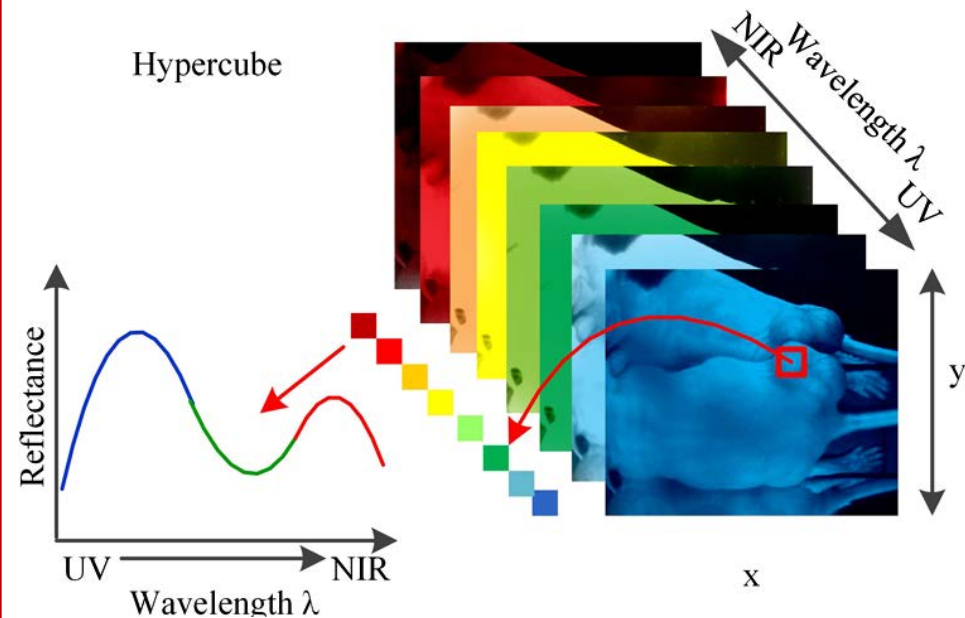


# Hyperspectral imaging

## RGB Imaging



## Hyperspectral Imaging

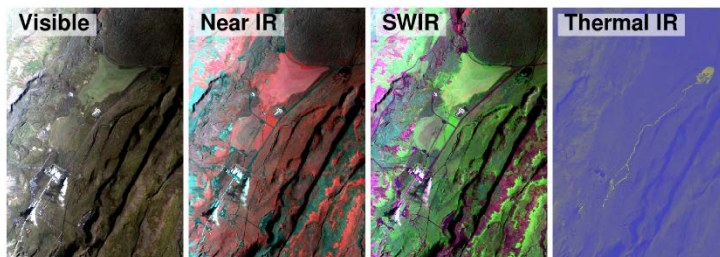


Hyperspectral imaging are based on the combined acquisition of an **image** and **spectral data**

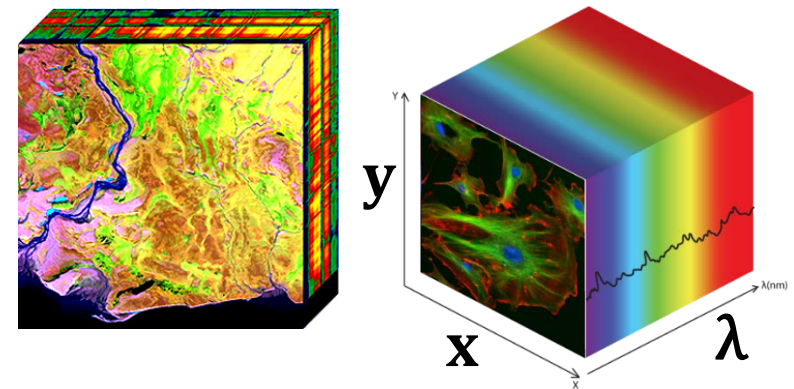
# Hyperspectral and multispectral imaging

- **Multispectral imaging** consists in acquiring an image within **narrow discrete spectral bands**
- **Hyperspectral imaging** provides one **full spectrum** for each pixel of the collected image

## *Multispectral imaging*



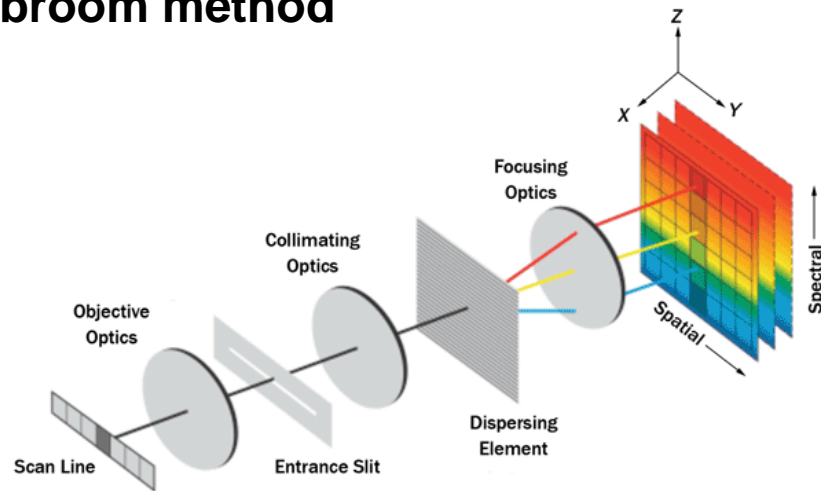
## *Hyperspectral imaging*



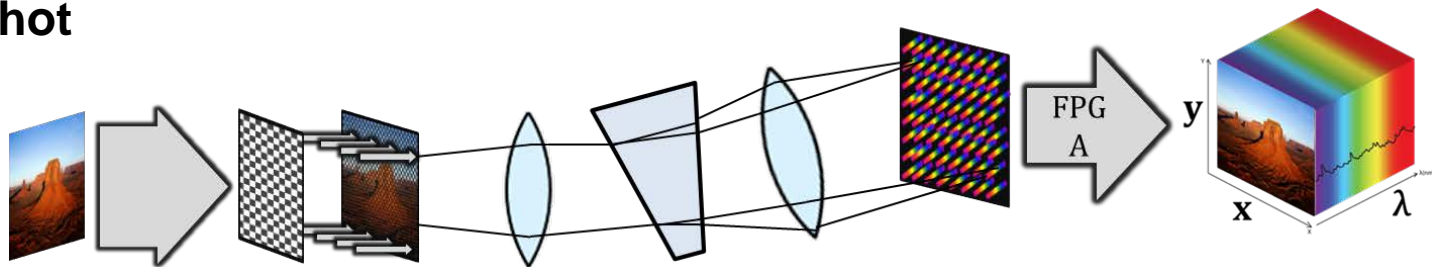


# Hyperspectral imaging acquisition technologies

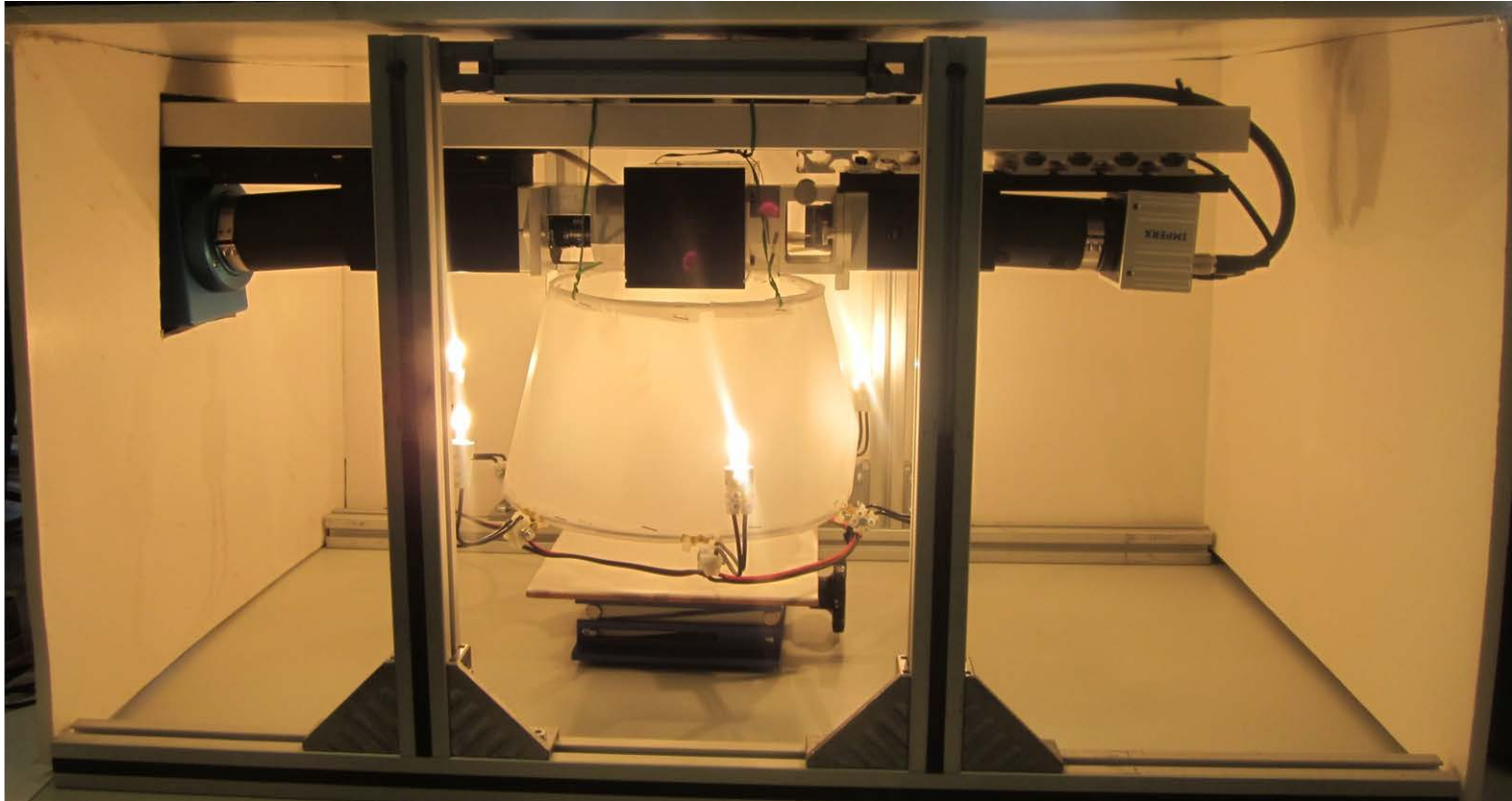
## 1. Scanning or push-broom method



## 2. Snapshot



# VIS and NIR hyperspectral cameras at the Televitis lab



- Visible: 400 - 1000 nm (1200 bands)
- Near Infrared: 900 - 1700 nm (256 bands)

# Hyperspectral imaging under field conditions

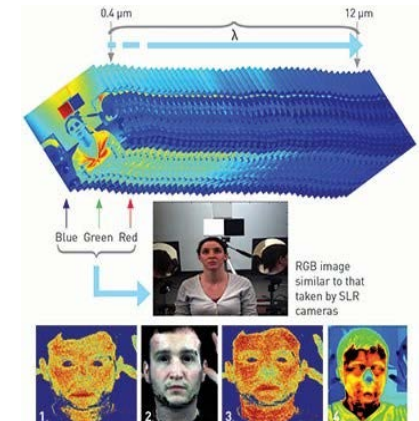
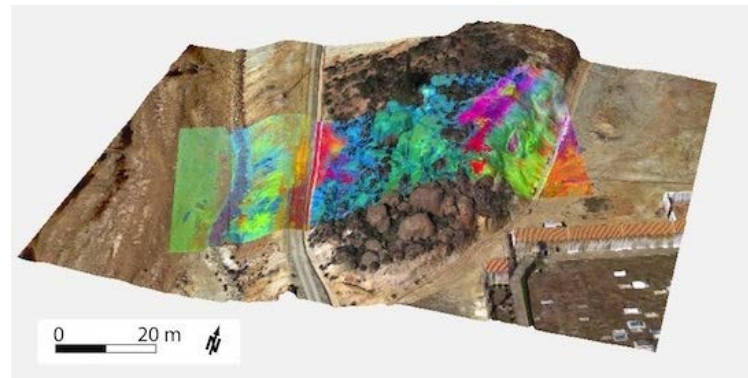
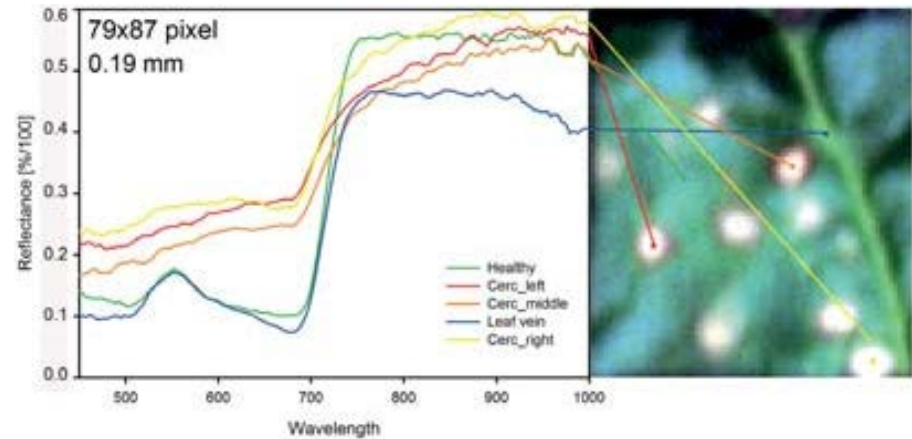


Tempranillo commercial vineyard  
Logroño, La Rioja, 2013

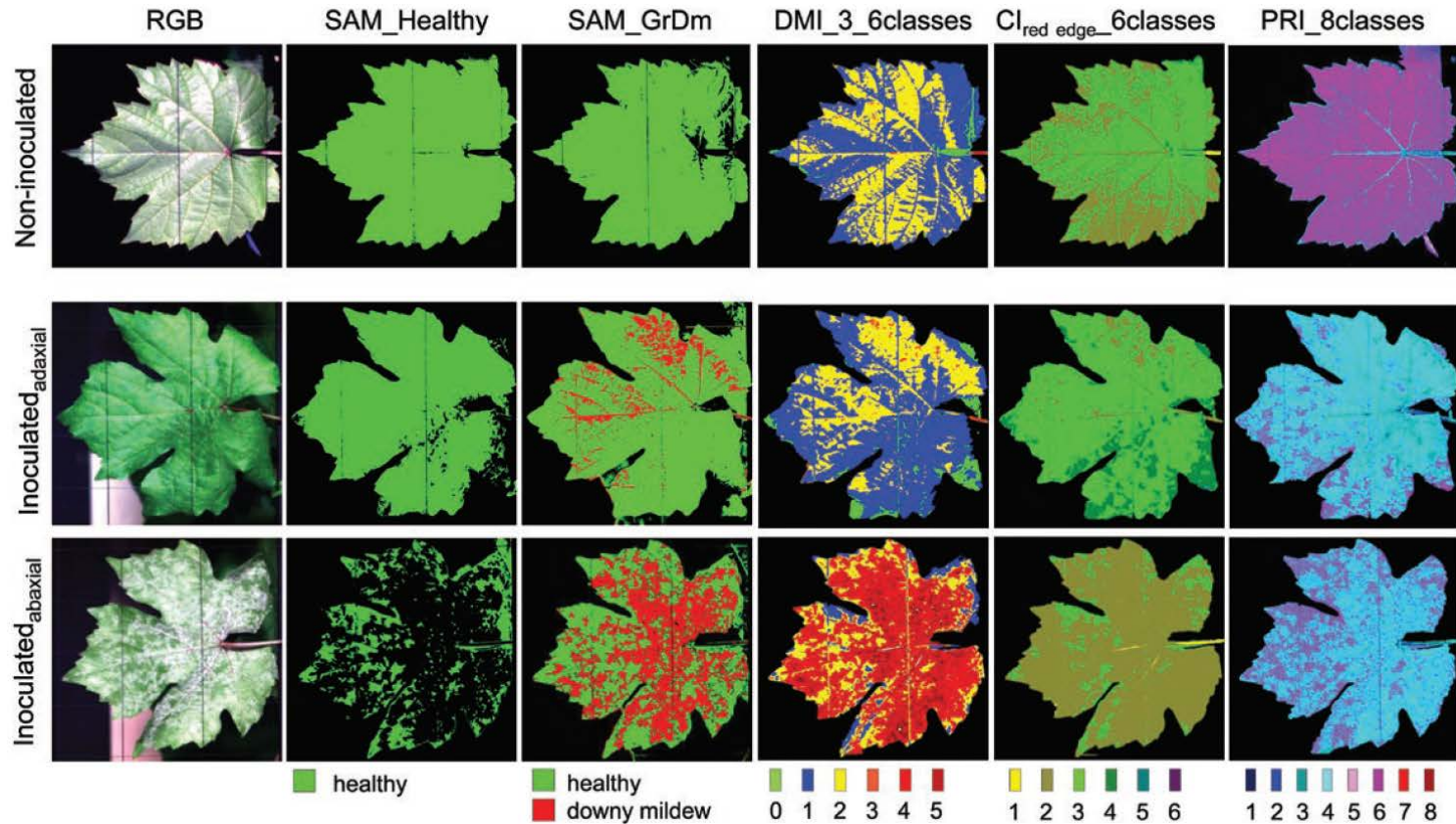


# Hyperspectral imaging applications

- Agriculture
- Forestry
- Environment
- Defence
- Medicine
- Water
- Food quality and safety control
- Geology
- Crime scene detection

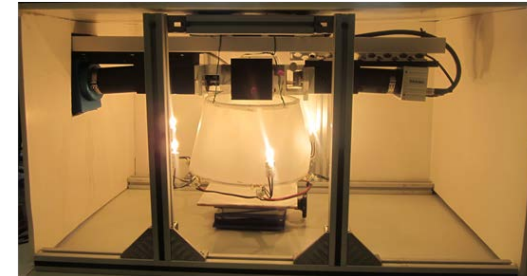
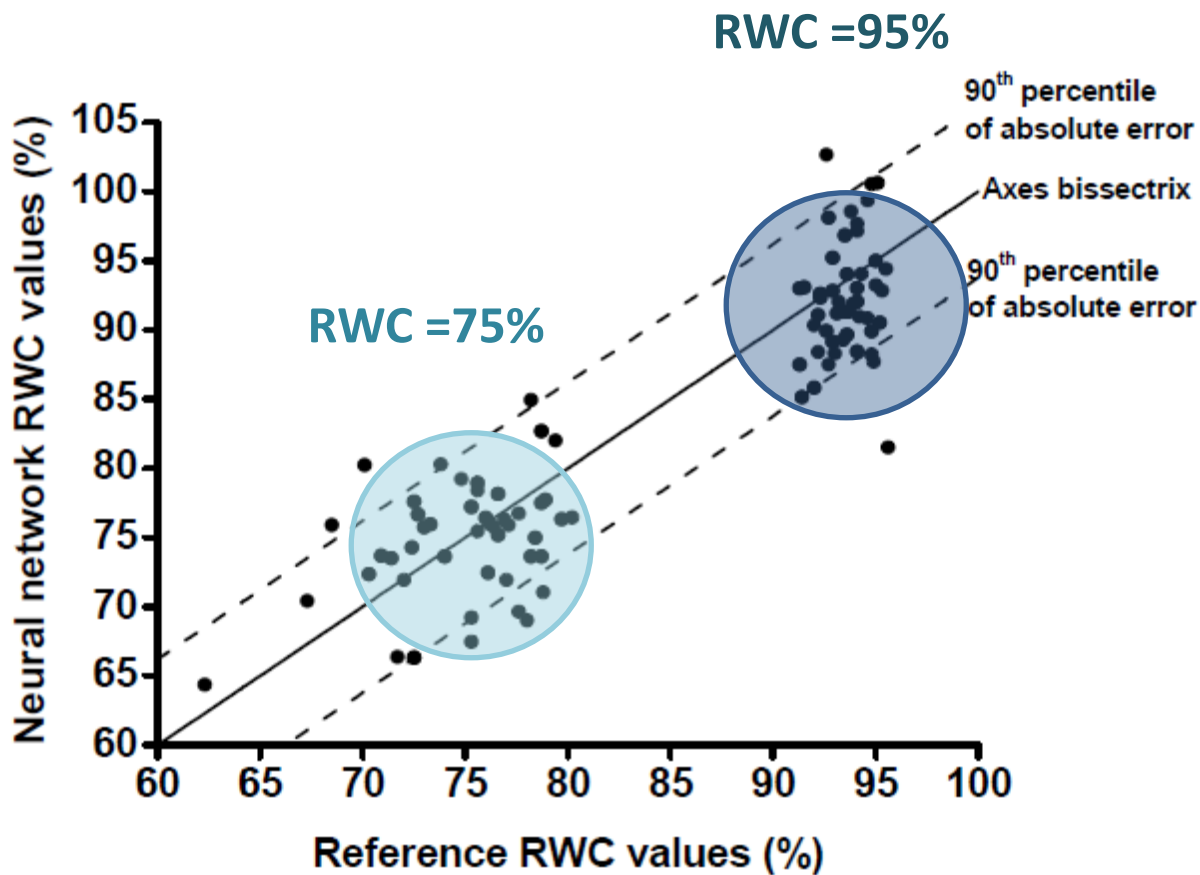


# Downey mildew phenotyping



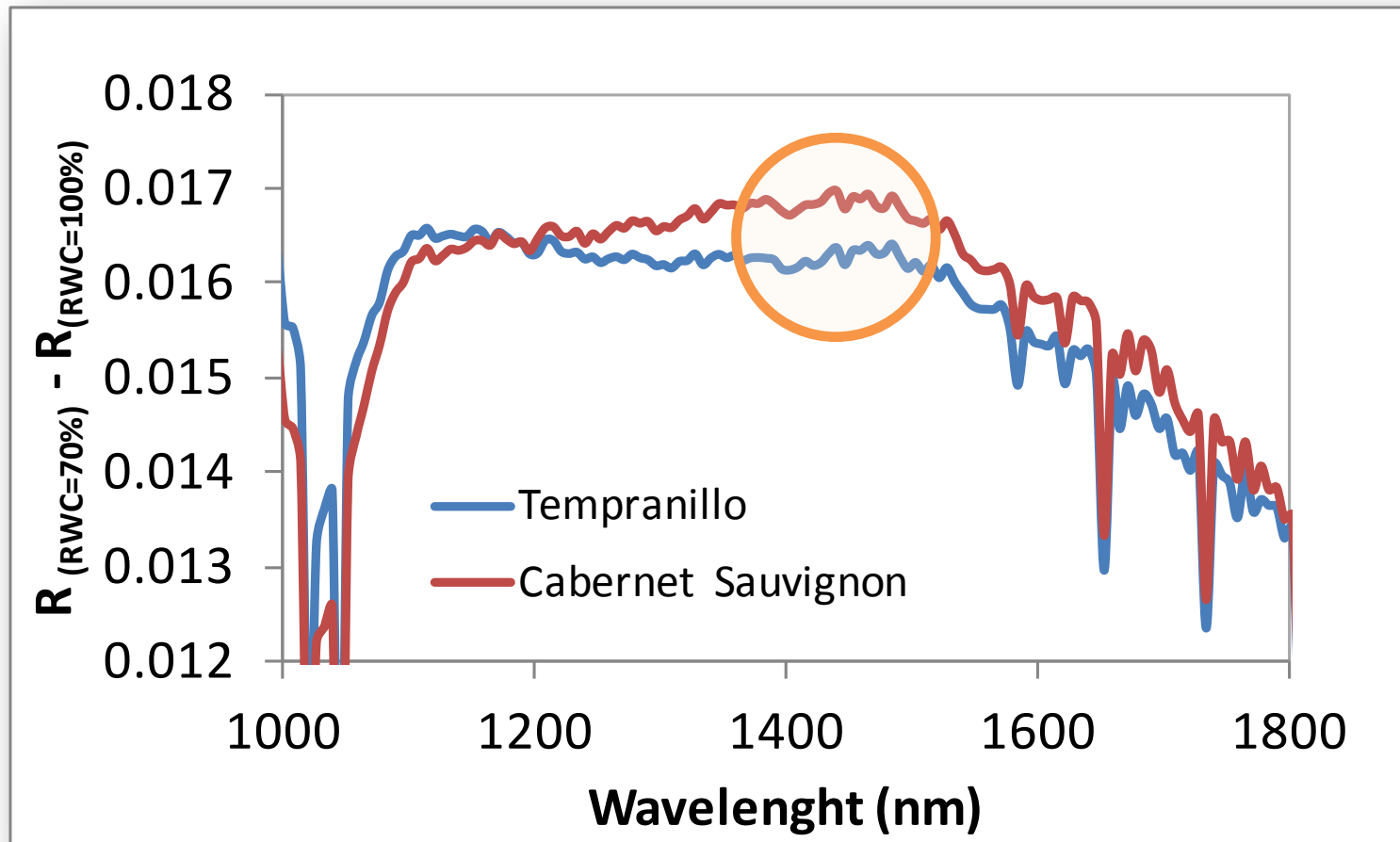
## Hyperspectral phenotyping of the reaction of grapevine genotypes to *Plasmopara viticola*

# Assessment of leaf water status



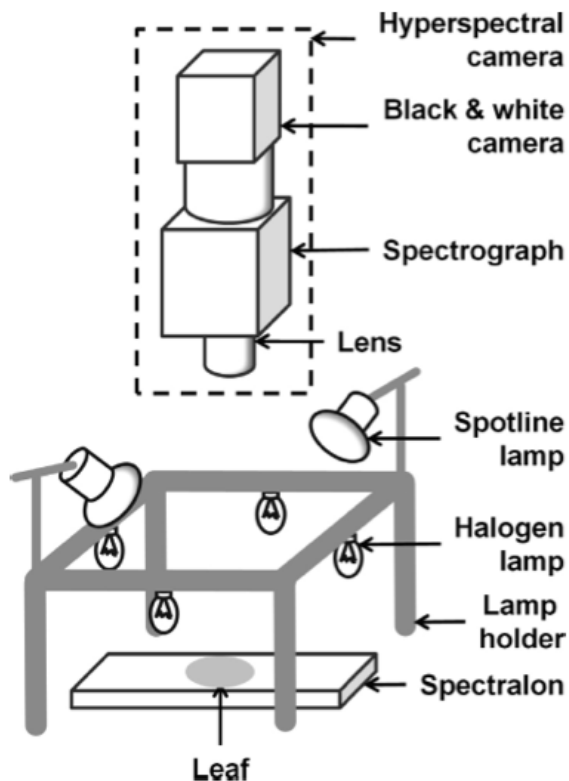
**Televitis**  
DATA-DRIVEN VITICULTURE™

# Spectral response to grapevine variety



# Discrimination of grapevine varieties and clones

**OBJECTIVE:** *Varieties and clones discrimination by hyperspectral imaging*



## IMPLEMENTATION:

### SPECTRA ADQUISITION:

- Passive sensor: hyperspectral cameras working in the visible and NIR range
- Using leaf disc images

### DATA MINING:

- Different preprocessing techniques: normalization
- PLS for classification
- Model evaluation: Montecarlo CV



# Classification of varieties and clones

Grenache



Clones

RJ 26

RJ11

ARA 4

ARA 24

Tempranillo



Clones

RJ 75

RJ 43

RJ 26

RJ 24

Cabernet Sauvignon



Clones

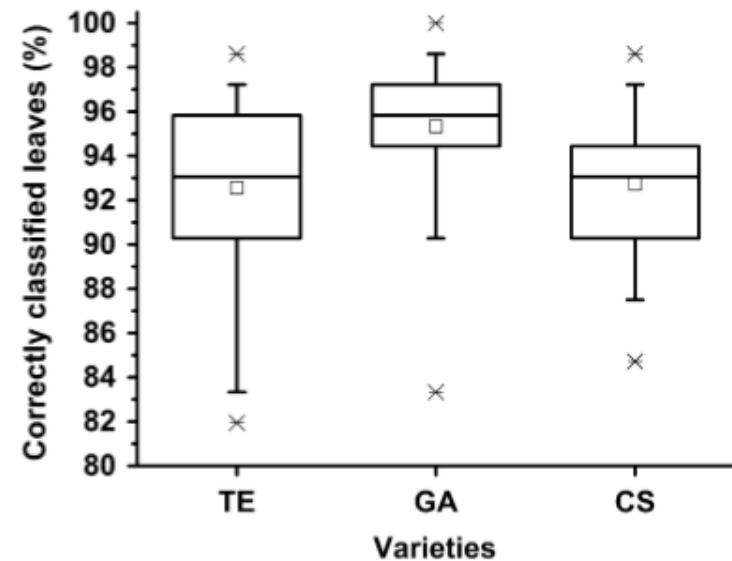
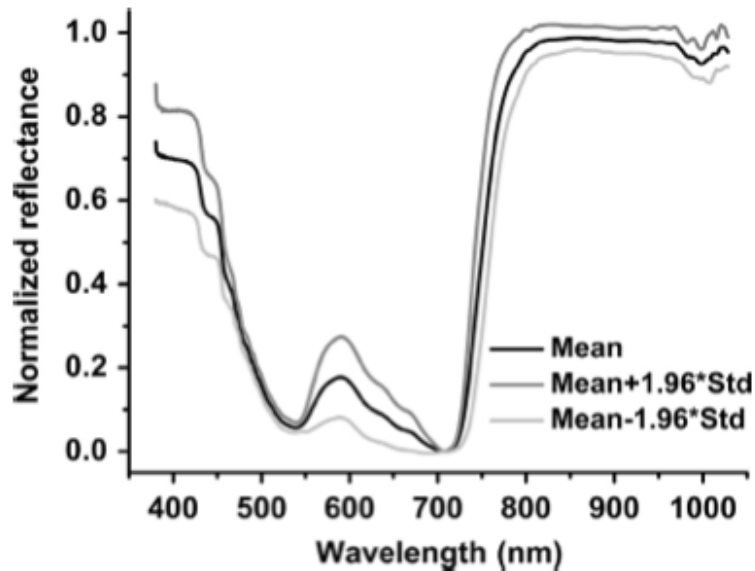
R5

169

15

685

# Classification of grapevine varieties

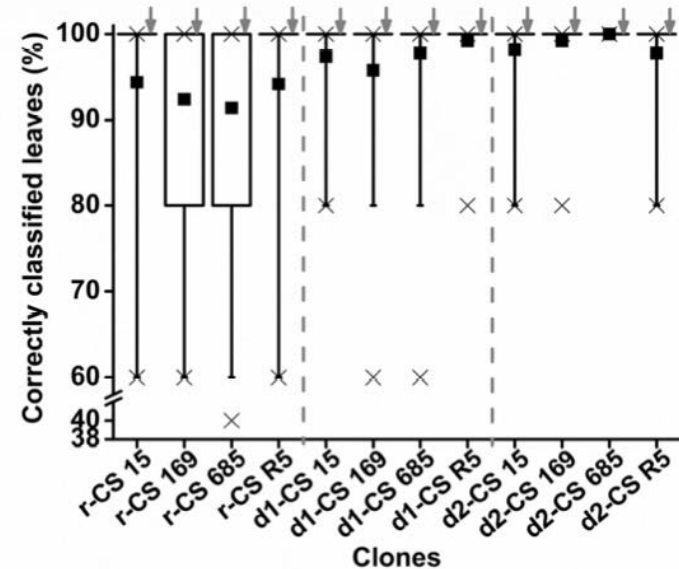
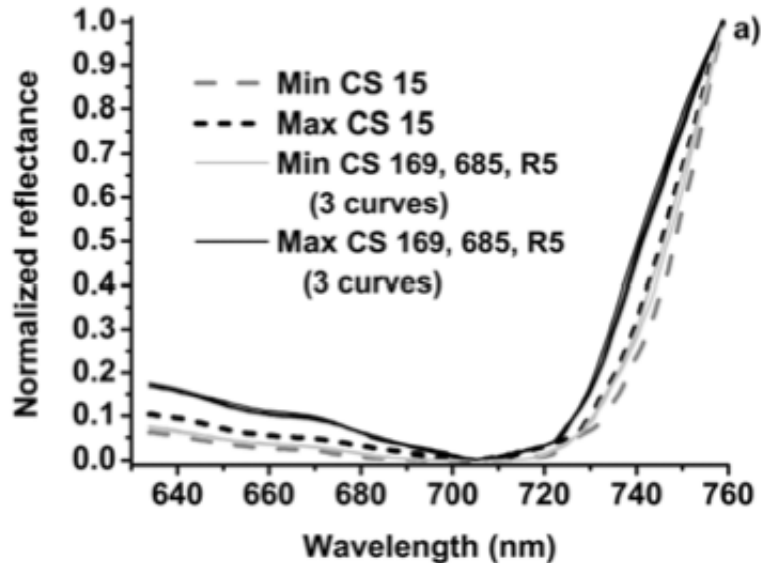


Diago, M.P., Fernandes, A.M., Millan, B., Tardaguila, J., Melo-Pinto, P 2013. Identification of grapevine varieties using leaf spectroscopy and partial least squares. *Computers and Electronics in Agriculture* 99, 7– 13.

Three correctly classified varieties:

- Grenache (GA)
- Cabernet Sauvignon (CS)
- Tempranillo (TE)

# Discrimination of grapevine clones



Fernandes, A.M., Melo-Pinto Millán, B., Tardáguila, J. Diago, M.P., 2015. Automatic discrimination of grapevine (*Vitis vinifera*) clones using leaves hyperspectral imaging and partial least squares regression. *Journal of Agricultural Science* 153:455-465

## Use of Visible and Short-Wave Near-Infrared Hyperspectral Imaging To Fingerprint Anthocyanins in Intact Grape Berries

Maria P. Diago,<sup>\*,†</sup> Juan Fernández-Navales,<sup>†</sup> Armando M. Fernandes,<sup>§</sup> Pedro Melo-Pinto,<sup>#,‡</sup>  
and Javier Tardaguila<sup>†</sup>

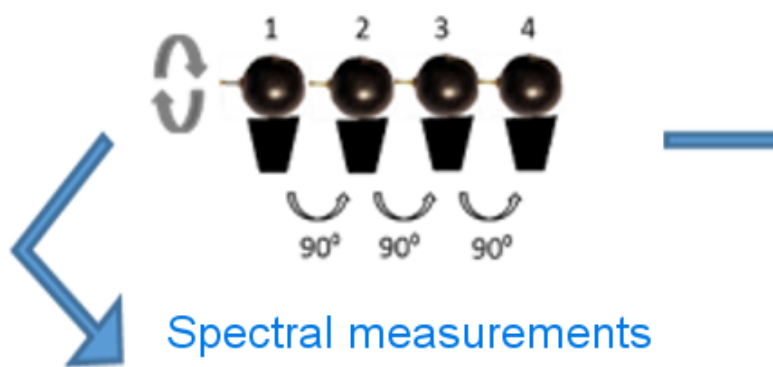
<sup>†</sup>Instituto de Ciencias de la Vid y del Vino (University of La Rioja, CSIC, Gobierno de La Rioja), Finca La Grajera, Ctra. Burgos Km. 6, 26007 Logroño, Spain

<sup>§</sup>INOV – INESC Inovação, Rua Alves Redol 9, 1000-029 Lisboa, Portugal

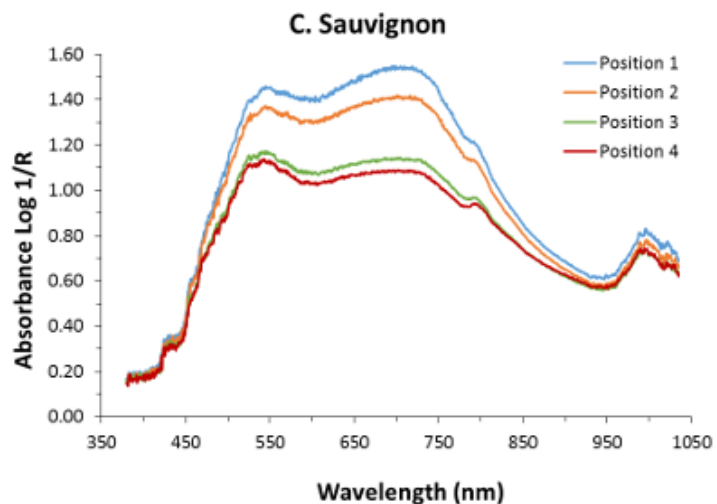
<sup>#</sup>CITAB-Centre for the Research and Technology of Agro-Environmental and Biological Sciences, Universidade de Trás-os-Montes e Alto Douro, Quinta de Prados, 5000-911 Vila Real, Portugal

<sup>‡</sup>Departamento de Engenharias, Universidade de Trás-os-Montes e Alto Douro, Quinta de Prados, 5000-911 Vila Real, Portugal

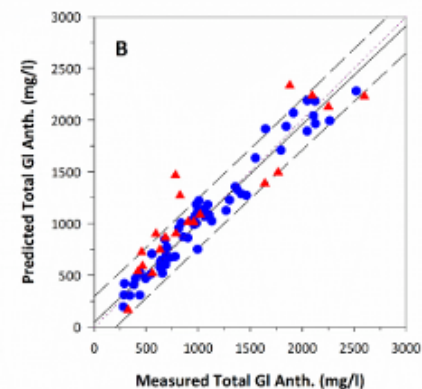
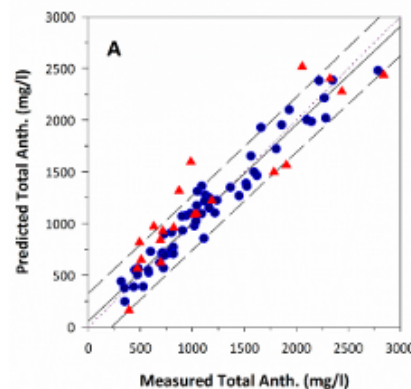
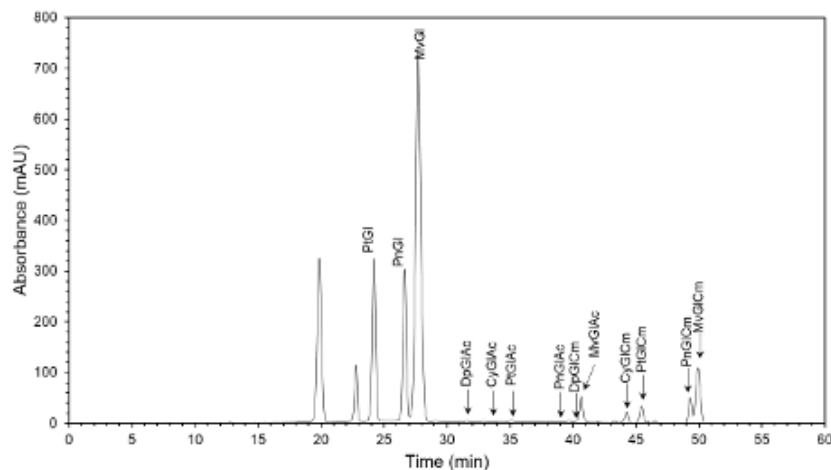
# Hyperspectral imaging for fingerprint anthocyanins



Spectral measurements

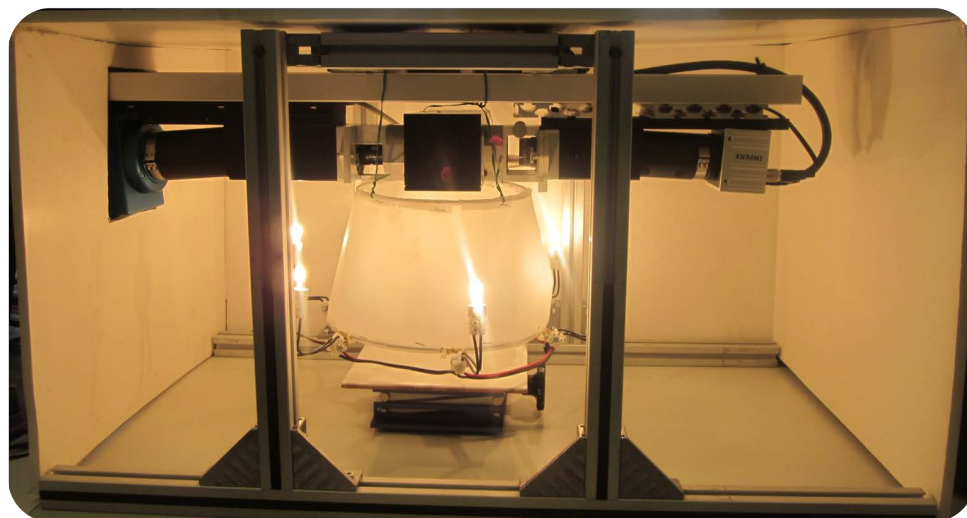


## Anthocyanin profile by HPLC-DAD

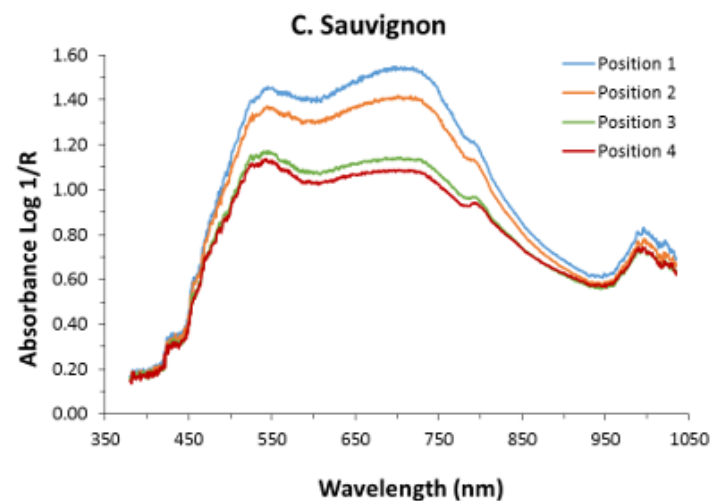
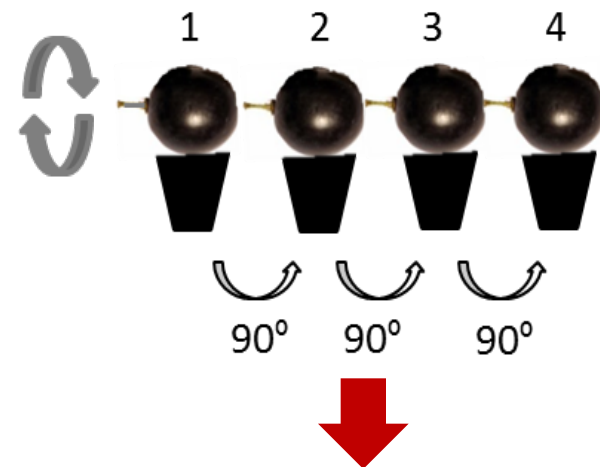


# Hyperspectral imaging to fingerprint anthocyanins

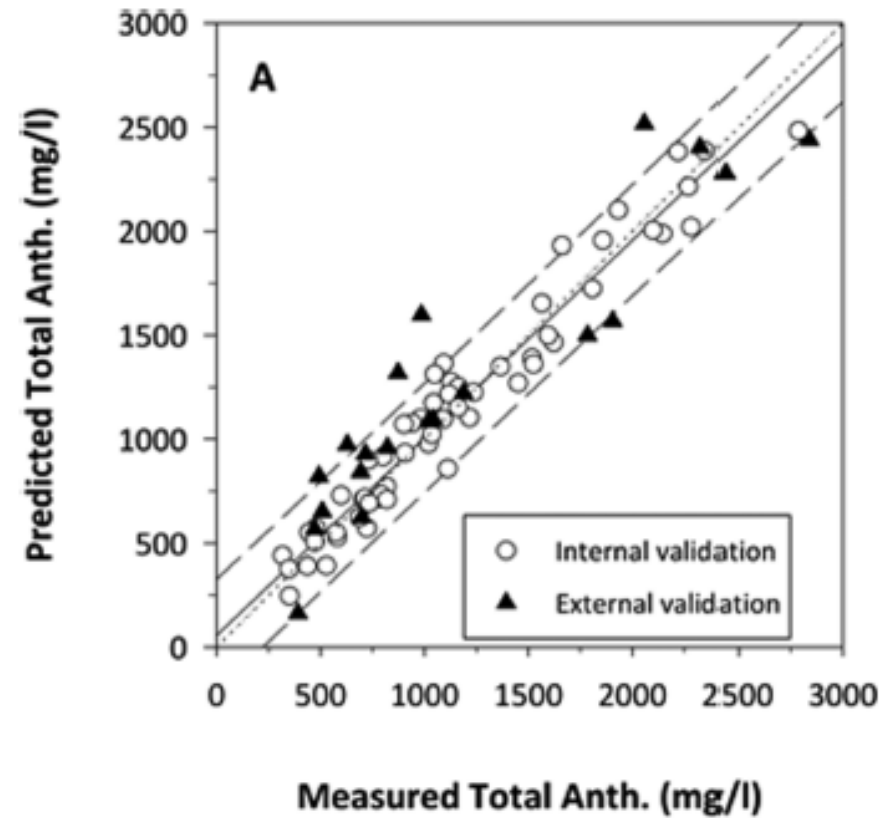
- 8 red varieties
- 10 berries/variety



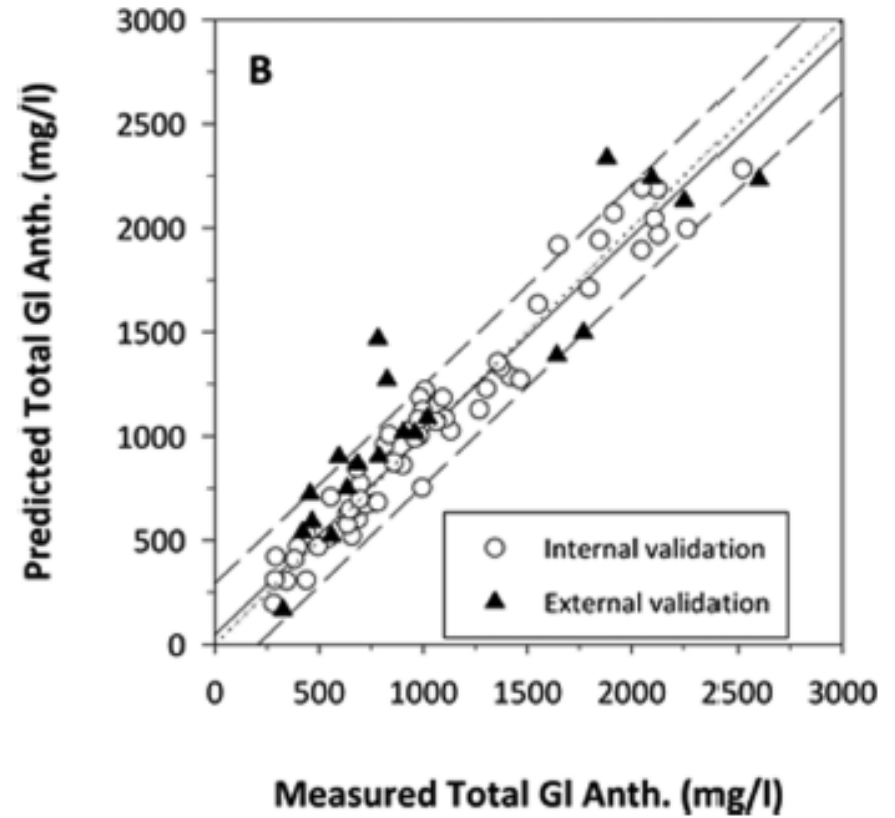
VIS-SWNIR Hyperspectral camera  
(400-1000 nm)



# Hyperspectral imaging to fingerprint anthocyanins



$$R^2_p=0.86$$



$$R^2_p=0.86$$

# Hyperspectral imaging to fingerprint anthocyanins

compound	spectral treatment	N	SD	minimum	maximum	PLS factor	calibration		cross-validation			external validation <sup>b</sup>	
							SEC	R <sup>2</sup> <sub>C</sub>	SECV	R <sup>2</sup> <sub>CV</sub>	RPD	SEP	R <sup>2</sup> <sub>P</sub>
Total Anth	Snv-DT 1.5.5.1	76	629.46	315.87	2839.96	4	148.25	0.94	189.05	0.91	3.51	281.58	0.86
Total Gl Anth	1.5.5.1	74	562.69	280.73	2521.24	4	125.97	0.95	155.94	0.92	4.03	272.60	0.86
Total GlAc Anth	2.5.5.1	73	12.80	1.16	54.26	5	1.98	0.98	4.12	0.90	4.10	4.82	0.51
Total GlCm Anth	1.5.5.1	73	56.49	4.02	221.37	6	13.42	0.94	23.09	0.83	2.55	42.72	0.40
MvGl	Snv-DT 1.5.5.1	76	201.85	226.56	1100.02	4	56.014	0.92	73.93	0.87	2.87	109.10	0.83
MvGlAc	2.5.5.1	48	13.39	0.09	46.49	6	1.32	0.99	4.13	0.90	4.27	3.30	0.90
MvGlCm	Snv-DT 1.5.5.1	75	29.78	0.73	119.94	7	6.99	0.94	13.32	0.80	2.33	23.33	0.57
PtGl	Snv-DT 1.5.5.1	75	108.00	16.25	426.94	4	23.29	0.95	29.44	0.93	4.01	49.66	0.87
PtGlAc	1.5.5.1	72	1.50	0.23	6.09	3	0.73	0.76	0.98	0.57	2.00	1.18	0.35
PtGlCm	1.5.5.1	71	6.72	0.28	26.06	7	1.12	0.97	2.04	0.91	3.98	4.44	0.84
DpGl	Snv-DT 1.5.5.1	78	158.65	9.67	657.71	4	36.89	0.95	48.41	0.91	3.78	71.39	0.88
DpGlAc													
DpGlCm	Snv-DT 1.5.5.1	72	11.59	0.50	46.49	7	1.83	0.97	3.28	0.92	4.51	2.34	0.48
PnGl	Snv-DT 1.5.5.1	73	86.47	10.06	383.09	7	15.93	0.97	30.35	0.88	3.46	49.10	0.81
PnGlAc	Snv-DT 2.5.5.1	69	0.69	0.02	2.82	2	0.26	0.86	0.31	0.80	2.61	0.35	0.88
PnGlCm	Snv-DT 2.5.5.1	71	7.56	0.34	37.53	4	2.14	0.92	3.81	0.75	2.38	6.50	0.54
CyGl	1.5.5.1	67	47.50	0.58	159.63	6	8.59	0.97	16.43	0.88	6.51	48.85	0.69
CyGlAc	1.5.5.1	54	0.33	0.01	1.26	2	0.14	0.81	0.16	0.77	3.94	0.16	0.85
CyGlCm	1.5.5.1	70	4.80	1.46	19.14	5	1.21	0.94	1.78	0.86	3.10	3.82	0.63

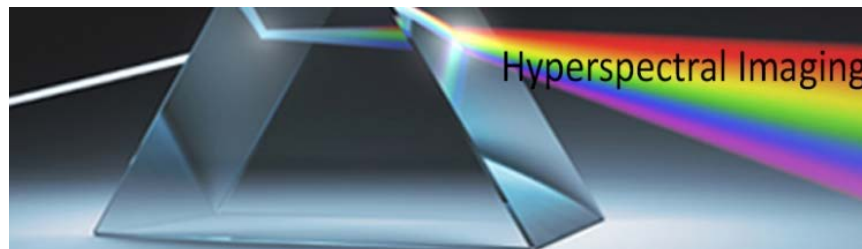


# Hyperspectral imaging to assess esca disease



# Conclusions

- ❑ Advanced and powerful technology for vineyard monitoring and phenotyping
- ❑ Indoor and outdoor working conditions
- ❑ Proximal and remote sensing applications
- ❑ Assessment of grape composition, water status, diseases, etc.
- ❑ High flexibility to target specific problems





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